United Arab Emirates University Scholarworks@UAEU

Theses

Electronic Theses and Dissertations

4-2022

EFFECT OF NATURAL ELICITORS AND COLD STORAGEPERIOD ON QUALITY IMPROVEMENT OF UAE DATE PALM FRUIT (PHOENIX DACTYLIFERA L. CV. KHESAB)

Shamsa Salem Nasser Abdulla Alblooshi

Follow this and additional works at: https://scholarworks.uaeu.ac.ae/all_theses

Part of the Agriculture Commons, and the Horticulture Commons



جامعة الإمارات العربيـة المتحدة (United Arab Emirates University



MASTER THESIS NO. 2022:9 College of Agriculture and Veterinary Medicine Department of Integrative Agriculture

EFFECT OF NATURAL ELICITORS AND COLD STORAGE PERIOD ON QUALITY IMPROVEMENT OF UAE DATE PALM FRUIT (PHOENIX DACTYLIFERA L. CV. KHESAB)

Shamsa Salem Nasser Abdulla Alblooshi



Enhanced nutritional quality

United Arab Emirates University

College of Agriculture and Veterinary Medicine

Department of Integrative Agriculture

EFFECT OF NATURAL ELICITORS AND COLD STORAGE PERIOD ON QUALITY IMPROVEMENT OF UAE DATE PALM FRUIT (*PHOENIX DACTYLIFERA* L. CV. KHESAB)

Shamsa Salem Nasser Abdulla Alblooshi

This thesis is submitted in partial fulfilment of the requirements for the degree of Master of Science in Horticulture

Under the Supervision of Dr. Zeinab Ahmed

April 2022

Declaration of Original Work

I, Shamsa Salem Nasser Abdulla Alblooshi, the undersigned, a graduate student at the United Arab Emirates University (UAEU), and the author of this dissertation entitled *"The Effect of Natural Elicitors and Cold Storage Period on Quality Improvement of UAE Date Palm Fruits (Phoenix dactylifera L., cv. Khesab)"* hereby, solemnly declare that this dissertation is the original research work that has been done and prepared by me under the supervision of Dr. Zeinab Ahmed, in the College of Agriculture and Veterinary Medicine at UAEU. This work has not previously been presented or published or formed the basis for the award of any academic degree, diploma or a similar title at this or any other university. Any materials borrowed from other sources (whether published or unpublished) and relied upon or included in my dissertation have been appropriately cited and acknowledged by appropriate academic conventions. I further declare that there is no potential conflict of interest concerning the research, data collection, authorship, presentation and publication of this dissertation.

Students Signature: Shamsa

Date: 23/05/2022

Copyright © 2022 Shamsa Salem Nasser Abdulla Alblooshi All Rights Reserved

Advisory Committee

- Advisor: Zeinab Ahmed
 Title: Assistant Professor
 Department of Integrative Agriculture
 College of Agriculture and Veterinary Medicine
- Member: Abdul Jaleel Cheruth
 Title: Associate professor
 Department of Integrative Agriculture
 College of Agriculture and Veterinary Medicine

Approval of the Master Thesis

The following Examining Committee Members have approved this Master's Thesis:

2) Advisor (Committee Chair): Zeinab AhmedTitle: Assistant Professor

Department of Integrative Agriculture

College of Agriculture and Veterinary Medicine

mit Signature

Date 23/05/2022

- 3) Member: Mohammed Abdul Mohsen Alyafei
- 4) Title: Associate Professor
 Department of Integrative Agriculture
 College of Agriculture and Veterinary Medicine

Ì

Date 23/05/2022

 Member (External Examiner): Zora Singh Title: Professor College of Science, Health, Engineering, & Education Institute: Murdoch University, Berth, Australia

work ph.

Signature

Signature

Date 23/05/2022

This Master Thesis is accepted by:

Dean of the College of Agriculture and Veterinary Medicine: Professor Bhanu Chowdhary

Signature:	Shamp Chardhan	Date:	06/06/2022
-			

Dean of the College of Graduate Studies: Professor Ali Al-Marzouqi

Signature:	Ali Hassan E	Date:	06/06/2022

Copy ____ of ____

Abstract

Date palm (Phoenix dactylifera L.) is one of the most important fruit trees grown in arid and semi-arid regions. Despite the immense capabilities of date palm, it is still a challenge to maintain fruit quality, marketability and shelf life. The objective of this study was to investigate the synergistic effect of a preharvest spray application of the natural elicitor chitosan (Ch) 1% alone and in combination with salicylic acid (SA) 2 mM and calcium chloride (Ca) 3%; (Ch, SA, Ca, Ch+Ca, Ch+ SA, Ch+ SA +Ca) on the quality parameters, shelf life and bioactive compound content of date fruit of the cultivar 'Khasab' during a cold storage period of 60 days. The results showed that all treatments significantly delayed fruit senescence/decomposition compared to the control. Fruits treated with Ch+ SA, followed by Ch and Ch+ SA +Ca had the least weight loss, colour change and decay after 60 days of storage. Fruit treated with Ch+Ca, SA, and Ca had significantly lower total soluble solids values and the highest phenolic, tannin, and flavonoid contents compared with control fruit. Antioxidant activities were observed in all treatments, with a significantly higher effect in Ch+ SA +Ca and Ch+ SA compared to the control. Our results show a synergistic effect of combining triggers to extend the shelf life of date fruit during cold storage by maintaining quality and reducing senescence/deterioration, and we recommend this as a promising strategy.

Keywords: Date Fruit, Natural Elicitors, Synergistic Effect, Salicylic Acid, Chitosan, Storage Quality, Shelf-Life, Marketability.

Title and Abstract (in Arabic)

تأثير بعض المحفزات الطبيعية والتخزين المبرد على تحسين جودة ثمار نخيل التمر صنف الخصاب في دولة الإمارات

الملخص

يعتبر نخيل التمر من اهم أشجار الفاكهة التي تنمو في المناطق الجافه وشبه الجافه. على الرغم من الإمكانيات الهائلة للنخيل، إلا أن الحفاظ على جودة الثمار الطازجة وقابليتها للتسويق وفترة صلاحيتها لا يزال يمثل تحديًا كبيرا. لذلك كان الهدف من هذه الدراسة تقييم التأثير التارزي لمعاملات الرش قبل الحصاد بمركب الكايتوسان الطبيعي 1/ (Ch) بمفرده أو بالخلط مع حمض الساليسيليك 2 ملى مول (SA) وكلوريد الكالسيوم (Ca) ٪3؛ أو خلط الثلاث ث مركبات معا (Ch + SA + Ca)، وذلك من خلال دراسة معايير جودة ومدة التخزين ومحتوى المركبات النشطة بيولوجيًا لثمار التمر من صنف "الخصاب" أثناء التخزين البارد لمدة 60 يومًا. أظهرت النتائج أن جميع المعاملات تؤخر بشكل معنوى شيخوخة الثمار وتدهورها مقارنة بالثمار الغير معاملة (الكونترول). الثمار المعاملة بالكايتوسان مع حمض الساليسيليك تليها الثمار المعالجة بالكايتوسان فقط ثم الفاكهة المعاملة بالكايتوسان مع حمض الساليسيليك وكلوريد الكالسيوم أظهرت انخفاض في فقد الوزن وتغير في اللون وأقل تدهور حتى بعد 60 يوم من التخزين البارد. بينما أظهرت الثمار المعاملة بالكايتوسان مع كلوريد الكالسيوم وكذلك المعاملة بحمض الساليسليك فقط أو بكلوريد الكالسيوم فقط أظهرت بشكل ملحوظ أقل تغير في المواد الصلبة الذائبة الكلية، وأعلى محتوى إجمالي للمركبات الفينولية والتانينات والفلافونيدات مقارنة بالثمار الغبر معاملة. أما بالنسبة الى النشاط المضاد للأكسدة وجد أن جميع المعاملات أظهرت نشاط لمضادات الأكسدة بالمقارنة بالكنترول. التأثير الأكبر كان ملحوظا على الثمار التي تم معاملتها بالكايتوسان وحمض الساليسيليك فقط والثمار التى تمت معاملتها بالكايتوسان مع حمض الساليسيلسك وكلوريد الكالسيوم. أظهرت نتائج هذه الدر إسة دليلًا على أن المحفز إت الطبيعية لها تأثير تآزري في إطالة عمر ثمار التمر أثناء التخزين البارد من خلال المحافظة على جودتها وتأخير الشيخوخة وتقليل التدهور. توضح النتائج أن استخدام المحفزات الطبيعية في تحسين جودة الثمار له نتائج واعدة وهي استراتيجية يوصى بها للاستخدام تجاريا على أصناف التمر الأخرى أو ثمار الفواكه الأخرى.

مفاهيم البحث الرئيسية: فاكهة التمر، المستخلصات الطبيعية، التأثير التارزي، حمض الساليسيليك، الكايتوسان، جودة التخزين، مدة الصلاحية، القابلية للتسويق

Acknowledgements

I would like to express my sincere gratitude to my supervisor first before anyone else, Dr. Zeinab Ahmed, Assistant Professor in the Department of Integrative Agriculture, UAEU, for her regular checking, encouragement, and guidance in every single step throughout this work until the very end of it, which made the work finish at ease. I would also like to thank the research assistant that worked with me Navjot Kaur for her help in the experiment and the explanation of the process. I am also thankful to my professors, the technical staff at the Department of Integrative Agriculture and the students who helped in the laboratory work for all their hard work to fulfil this work. My sincere thanks extended to Ahmed Taha (Library Research Desk) for providing an expert editing service. Finally, I would like to thank my family for their continuous support from the very beginning, without all of you it would not be possible.

Dedication

To my country the United Arab Emirates, my young self that did not expect me to go this far, my parents, my siblings God's greatest blessing, my teachers, instructors, and professors. Thank you for believing in me even when I could not. You made me who I am, and I am forever grateful for that.

Table of Contents

Title	i
Declaration of Original Work	ii
Copyright	iii
Advisory Committee	iv
Approval of the Master Thesis	
Abstract	vii
Title and Abstract (in Arabic)	viii
Acknowledgements	
Dedication	
Table of Contents	
List of Tables	
List of Figures	
Chapter 1: Introduction	
1.1 Date Palm	
1.2 Date Fruit	1
1.2.1 Fruit growth, development and maturity stages	3
1.2.2 Nutritional value and health benefits of dates	5
1.3 Date Palm Cultivars	7
1.3.1 Khesab cultivar	7
1.4 Use of Natural Elicitors	8
1.5 Date palm Fruits in the UAE	10
1.6 Research Objectives	11
Chapter 2: Literature Review	12
2.1 Introduction	12
2.2 Chitosan as Natural Elicitors	12
2.2.1 Preharvest application of chitosan	13
2.2.2 Postharvest applications of chitosan and their derivatives	15
2.2.3 Treatments with chitosan and combination of calcium chloride	
and salicylic acid	18
2.3 Current Strategies for the Preharvest/Post-Harvest Treatment of	
Date Fruit	24
Chapter 3: Materials and methods	26
3.1 Plant Material and Sampling	26
3.2 Physiochemical analysis	27
3.2.1 Fruit characteristics	27

3.2.2 Fruit ripening and decay percentage	
3.2.3 Total soluble solids (TSS)	
3.2.4 Fruit surface colour	
3.2.5 Determination of total microbial load	29
3.2.6 Extraction of bioactive compounds	29
3.3 Phytochemical Analysis	29
3.3.1 Total Phenolic Content (TPC)	29
3.3.2 Total Tannin Content (TTC)	30
3.3.3 Total Flavonoids Content (TFC)	30
3.4 Antioxidant Activities	31
3.4.1 DPPH radical scavenging activity	31
3.4.2 ABTS radical scavenging activity	31
3.5 Statistical Analysis	32
Chapter 4: Results and Discussion	33
4.1 Physical Quality Characteristics of Fruit at Harvest	33
4.2 Fruit Weight Loss during Storage	33
4.3 Total Soluble Solids (TSS) during cold storage	36
4.4 Fruit Ripening	38
4.5 Fruit Decay During Storage	41
4.6 Colour Attributes	43
4.7 Total Phenolic Content of Fruit at Harvest and During Storage	45
4.8 Total Tannin, Flavonoids, and Antioxidants Concentrations at	
Harvest	48
4.9 Microbiological Quality of Date Fruit	51
Chapter 5: Summary and Conclusions	54
References	55

xii

List of Tables

Table 1: Physiological changes in fresh fruits after chitosan treatment and	
with other elicitors	20
Table 2: Preharvest spry treatments of Khesab cv. date fruit with different	
elicitors	27
Table 3: potential effects of different preharvest treatments on Khesab cv	35
Table 4: Effect of different preharvest treatments on Khesab cv. fruit colour	
attributes during cold storage	47
Table 5: Pearson's correlation coefficients between some biochemical and	
physical characteristics of Khesab cv. fruit	48
Table 6: Effect of different preharvest treatments on Khesab cv. fruit phyto-	
chemical analysis at harvest	51

List of Figures

Figure 1: Major date producing countries in the world	2
Figure 2: Major date fruit exporter in the world	3
Figure 3: The Bisr and half Rutab fruit maturity stages of Khesab date fruit	5
Figure 4: Effect of different preharvest treatments on weight loss of Khesab	
cv. fruit during cold storage	34
Figure 5: Effect of different preharvest treatments of Khesab cv. fruit during	
cold storage	38
Figure 6: Effect of different preharvest treatments on Khesab cv. fruit ripening	
during cold storage	40
Figure 7: Effect of different preharvest treatments on Khesab cv. fruit decay	
during cold storage	42
Figure 8: Effect of different preharvest treatments on total phenol content of	
Khesab cv. fruit during cold storage	43
Figure 9: Effect of preharvest treatment on viable microbial load (Log10	
CFU/g) of Khesab cv. at day 0 and day 60 of cold storage	53

Chapter 1: Introduction

1.1 Date Palm

Date palm (*Phoenix dactylifera* L.) is one of the most important fruit trees grown in arid and semi-arid regions. The palm family (*Arecaceae*) is comprised of 183 genera and over 2,400 species. Five major palm species are domesticated fully and are grown as economic species, date palm (*Phoenix dactylifera*) is one of them. Date palm is a diploid (2n=36), perennial and monocotyledonous (Haider et al., 2013). The tree thrives in the hot and dry regions of Middle Eastern and North African countries and is highly valued as a 'blessed tree' known since ancient times. It is an essential part of the cultural heritage, traditional books, and poems of Arabian countries. The date palm substantially contributes to the economic development of these countries (Gantait et al., 2018; Mattar et al., 2021).

Despite the changes throughout the years, dates continue to play an essential role in the region (Ahmed et al., 1995). The date palm has intangible value in desert climates including creating shade and microclimatic conditions suitable for the growth of other crop plants along with functioning as windbreaks and stabilizing earthen irrigation works (Johnson, 2012). In other areas that have different climates including the Americas, southern Europe, Asia, Africa and Oceania, date palm is grown as an ornamental plant (Dransfield et al., 2005).

1.2 Date Fruit

Date palm tree usually at the age of 5 years produces 400–600 kg of date fruit per tree in a year and the fruiting continues up to 60 years. The fruit are arranged on spikelets bearing 20-60 individual dates, and several such spikelets are attached to a central stalk to form a bunch, each tree has between 5 to 30 brunches (Ahmed et al., 1995). Date fruit can be sold fresh or dried, and they can be stored well under low-temperature conditions (Johnson, 2012). The consistency of the fruit varies from soft to dry and the colour varies from yellow to black or from red\dark pink to black depending on the type of the cultivar. Each fruit is a one-seeded berry consisting of a fleshy mesocarp covered by a thin epicarp and the seed is ventrally grooved with a hard endosperm made of a cellulose deposit and usually oblong with a small embryo.

The characteristics of the date fruit and seed can vary depending on the environmental conditions, field management and the cultivar itself. (Ahmed et al., 1995; Zaid & de Wet, 2002; Al-Yahyai & Kharusi, 2012, Ahmed et al., 2022a). When it comes to date fruit production, the top ten producing countries around the world are Egypt, Saudi Arabia, Iran, Algeria, Iraq, Pakistan, Sudan, Oman, Tunisia and the United Arab Emirates, as shown in Figure 1.

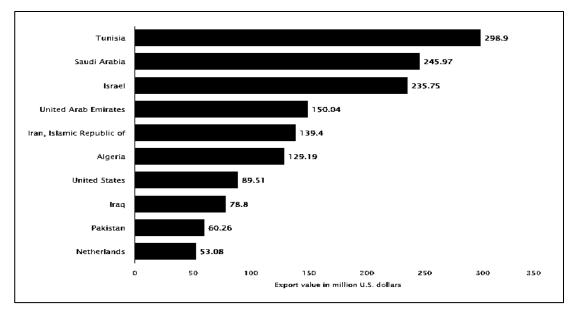


Figure 1: Major date producing countries in the world (FAO, 2020)

On the other hand, top ten exporting countries around the world are Tunisia, Saudi Arabia, UAE, Iran, Algeria, the USA, Iraq, Pakistan, and Netherlands, as shown Figure 2.

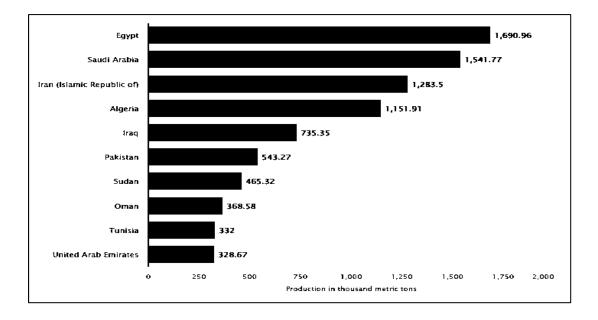


Figure 2: Major date fruit exporter in the world (FAO, 2020)

1.2.1 Fruit growth, development and maturity stages

The maturity of the date fruit varies depending on the cultivar, some of them mature early in the season and the others do not mature until the end of the season. the mature fruit can vary widely in its organoleptic, physical, and chemical characteristics (Ahmed et al., 1995). yet, all the edible fruits go through the same five stages which under the Arabic terminology, Hababouk, Kimri, Bisir or Khalal, Rutab and Tamer in that particular order (Awad et al., 2011a). These stages characterize the cell division, immature green (cell elongation), mature hard full coloured, soft brown, and firm raisin-like fruit, respectively (Awad et al., 2011b). In general, the harvesting and marketability of the date fruit depend on the cultivar in the context of the level of astringency, physiological conditions, and public demands, for this reason, they are delivered to the market at three stages which are: Bisr (full size, crunchy, coloured and mostly edible), Rutab (the fruit apex becomes soft and brown, most cultivars consumed fresh at this stage) and Tamer (hazel to dark brown, wrinkled, raisin-like dry fruit) (Mohamed et al., 2014).

After fertilisation, the *Hababouk* stage comes, and it is the stage where the loss of two unfertilized carpels occurs. The colour of the date fruit in this stage is creamy to light green and the fruit is not edible yet. Following the *Hababouk* stage, the Kimri stage comes, which is the green stage which lasts about 9 weeks depending on cultivar and location. The date fruit in the Kimri stage is having high water content. The third stage is Bisir which is also known as Khalal, it lasts about 4 to 5 weeks. The fruit attains physiological maturity with a slight decrease in size, weight, and starch content at Khalal stage (Al-Mssallem et al., 2013). In the Bisir stage the colour of the date fruit change from green to either yellow, pink, or red, or yellow spotted with red depending on the cultivar.

The fourth stage is Rutab, and it is the stage where the date fruit starts to accumulate reducing sugars and losing weight. At this stage, the fruit becomes softer, and the colour turns to light brown. The fruit progressively loses water (30–45%), and starch is converted to sugars during the Khalal and Rutab stages (Figure 3). The fifth and the last stage of the date fruit maturation is Tamer. In the Tamer stage, the date fruit is very sweet, its colour is dark brown, and its texture is soft with very low moisture content. The very low moisture content of the fruit in the Tamer stage leads to it is

ideal for long-term storage to be consumed during off-season (Kader and Hussein, 2009).



Figure 3: The Bisr and half Rutab fruit maturity stages of Khesab date fruit

1.2.2 Nutritional value and health benefits of dates

Apart from the agricultural and commercial importance of the date fruit in the Arabic Gulf region, the fruit has high nutritional value including dietary fibres (6.4-11.5%), carbohydrates (total sugars, 44-88%), proteins (2.3-5.6%), fat (0.2-9.3%), vitamins, minerals, phenolic, tannins, and antioxidants compounds (El Hadrami and El Hadrami, 2009; Vayalil, 2013; Al-Alawi et al., 2017; Al-Mssallem et al., 2020; Bentrad and Hamida, 2020). A study in the UAE showed that the date is an important source of potassium for regular consumers, and it stated that it provides the Arab world with a remarkable dietary component (Ahmed et al., 1995). In addition, date fruits are a good source of vitamin B complex, such as thiamine (B1), riboflavin (B2), niacin (B3), pantothenic (B5), pyridoxine (B6), and folate (B9) (Chao and Krueger, 2007; Al-Harrasi et al., 2014). Depending on many factors including the soil type, agronomic

practices, cultivar type and fruit maturation stage, the nutritional composition can differ (Al-Farsi et al., 2007; Amira et al., 2012).

Date flesh is a readily available source of energy, and this is because of the availability of high content of reducing sugars such as glucose and fructose (70-85% fresh weight) (Al-Farsi & Lee, 2008; Rastegar et al., 2012). The presence of non-enzymatic antioxidants (phenolic, flavonoid and ascorbic acid) and enzymatic antioxidants (catalase, peroxidase and superoxide dismutase), compounds enhance the free radical scavenging activity and therapeutic effects of dates (Biglari et al., 2008; Mohamed et al., 2014). Studies on date fruit showed that they can be beneficial when they are being consumed, where the polyphenolics account for the broad range of biological effects of dates among the other dietary components that the fruit include. This biological effects includes antimutagenic and antioxidant properties (Vayalil, 2002; Duthie et al., 2003).

The dietary fibres in dates also modulate the immune system and play a major role in the prevention of cardiovascular diseases (Frankel et al., 1993). Dates have anti-viral, anti-bacterial and anti-fungal properties which inhibit chronic inflammations, and this is because of the presence of phenolics (Kaul & Khanduja, 1998). The aqueous and ethanolic extracts of dates are effective in improving the severity of gastric ulceration (Al-Qarawi et al., 2003). Other human health benefits come from the hydroxycinnamates and flavonoids including tannins which are famous for being beneficial against cancer and cardiovascular diseases (Biglari et al., 2008).

1.3 Date Palm Cultivars

Date palm holds the distinction of having the greatest number of named cultivars based on the fruit characteristics among the other types of palms. Date fruit overwhelmingly represent the economic value of the tree (Johnson, 2012). Depending on the flesh consistency and moisture content at the fully-ripen stage, date palm cultivars can be divided into three groups which are namely soft, semi-dry and dry (Yahia & Kader, 2011). The major soft cultivars are *Hillawi, Abada, Amhat, Barhi, Bentaisha, Halawy, Hayani, Honey, Khadrawy*, and *Medjhool*.

During ripening, sucrose is converted into invert or reducing sugars (glucose and fructose) in soft cultivars with a moisture content greater than 30%. Cultivars such as *Badrayah, Bartamoda, Deglet Beida, Horra, Sakoty*, and *Thoory* are dry date cultivars with <20% moisture. Cultivars with 20–30% moisture are the semi dry-date cultivars which include *Amry, Dayri, Deglet Nour, Khalas, Sewy,* and *Zahidi*. In addition to reducing sugars, both dry and semi-dry dates retain a good amount of sucrose on full ripening (Chao and Krueger, 2007). Date palm cultivars are different in fruit characteristics, size and sugar content and this leads to more classifications that can be found within the same group.

1.3.1 Khesab cultivar

Khesab cv. is considered a late-maturity type of date palm cultivar. it has a high productivity character, and it can be consumed in the Rutab and tamer maturity stage. The colour of Khesab cv. varies depending on the stage of maturity. In the Bisir stage, the colour is red, brownish-red in the Rutab stage and dark brown when it is in the Tamer stage. The shape of the fruit is oval and oblong, the ratio of the weight of the fleshy part to the seed is large (Ahmed et al., 2021a). The flowering of Khesab cv. is at the end of February and the harvesting process take a place in the middle of August. Khesab cv. is resistant to salinity and moisture, it is also slightly frost-resistant which means that it can be stored in low-temperature conditions.

Khesab cv. is a quite common cultivar in the UAE that is preferred to be consumed at the half or full Rutab stages (Figure 3). However, upon harvesting, the fruit starts losing its firmness, colour, taste, and overall physiology which eventually lowers its market value (Al-Qurashi & Awad, 2011). These changes are outcomes of the postharvest activity of various ripening related enzymes and hormones that soften the fruit tissue, decrease the firmness and actual integrity of the fruit, and ultimately lead to the fruit's senescence (Abu-Shama et al., 2020). Therefore, slowing down the ripening process and senescence is critical in terms of commercial advantages (Mohamed et al., 2014). It is important to hold the date fruits in cold storage at a fully mature stage to extend their shelf life of it (Abd Elwahab et al., 2019 Ahmed et al., 2021a). In recent years, cold storage of date fruits has received more attention in the major date producing countries. Date industries usually store dates at 3°C for up to a year (Ismail et al., 2008; Al-Yahayai & Al-Kharusi, 2012).

1.4 Use of Natural Elicitors

The application of natural elicitors is used to enhance physiological adaptations and accelerate the plant's c defence system (Ahmed and Palta 2010; Baenas, 2014; Moreno-Escamilla et al., 2018). Many different studies on pre and postharvest applications have gained success in different fruit crops by using eco-friendly elicitors such as chitosan (Romanazzi et al., 2013; Shen & Yang, 2017; Rahman et al., 2018; Romanazzi et al., 2020), salicylic acid (Zhang et al., 2003;

Sayyari et al., 2009; Luo et al., 2011; Mohamed et al., 2014; Chen et al., 2020; Gomes et al., 2021), and calcium chloride (Irfan et al., 2013; Sohail et al., 2015; Atia et al., 2018), retarding tissue softening, delaying ripening, enhancing quality, extending shelf life, and preventing fruit deterioration.

Chitosan is one of the most recognized natural elicitors known for its antimicrobial, eliciting, and film-forming characteristics which is widely used in agriculture as a soil modifier, coating films, fungicide and elicitor (Romanazzi et al., 2013; Katiyar et al., 2014; Moreno-Escamilla et al., 2018; Rahman et al., 2018; Romanazzi et al., 2018, Ahmed et al., 2021a). It is a partially deacetylated derivative obtained from the chitin of crustacean shell wastes which is considered the second most abundant and important natural biopolymer in the world after cellulose (Hassan & Chang, 2017). Chitosan can control plant diseases and it induce resistance in the host plants and enhance biodiversity in the rhizosphere (Hassan & Chang, 2017). Pre- and post-harvest studies in different fruit crops have shown the positive effects of chitosan on the quality and the shelf and storage life of fruit and vegetables (Romanazzi et al., 2013; Shen & Yang, 2017; Romanazzi et al., 2018; Rahman et al., 2018, Ahmed et al., 2021b).

Salicylic acid (SA) is a simple phenolic phytohormone with various roles in plant growth and developmental processes. It also shows great potential for preventing the deterioration of fruits and vegetables postharvest (Shen & Yang, 2017, Ahmed et al., 2022b; Khalil et al., 2022). Salicylic acid can dramatically enhance disease resistance in mango fruit. In addition to that, studies stated that pre-harvest and post-harvest treatments of salicylic acid improved the quality and shelf life of table grapes (Pila et al., 2010). Salicylic acid preserves firmness reduces weight loss, colour progress, and disease incidence in plums, strawberries, peach and pears fruits for up to five weeks without decay (Khadiga, 1993).

Calcium chloride is another natural elicitor that has a well-known role in the physiology of plant tissue. It is also considered the most important mineral element which determines fruit quality (El Badawy, 2012). The use of calcium chloride after the fruit has been harvested maintains cell turgor, membrane integrity, tissue firmness, and delays membrane lipid catabolism (Atia et al., 2018). Calcium improves the rigidity of cell walls and obstructs enzymes such as polygalacturonate from reaching their active sites, thereby retarding tissue softening and delaying ripening. The use of calcium chloride after the fruit has been harvested maintains cell turgor, membrane integrity, tissue firmness and delays membrane lipid catabolism thus extending the storage life of fresh fruits (Pila et al., 2010).

These elicitors exert their effects by employing different mechanisms such as triggering the synthesis of phytochemicals, enhancing the production of particular antioxidant enzymes, and reducing ethylene production in vegetables and fruit (Ruiz-García & Gómez-Plaza, 2013; Baenas et al., 2014; Romanazzi et al., 2018). The active amino and hydroxyl groups of chitosan tend to react with the carbonyl group of salicylic acid. Hence, introducing salicylic acid into the backbone chain of chitosan can induce the formation of a chitosan-g-salicylic acid (CTS-g-SA) conjugate, which has excellent antibacterial effects and water solubility (He et al., 2011).

1.5 Date palm Fruits in the UAE

In the UAE, the date palm is a very remarkable and precious tree, which has strong religious, traditional, and nutritional significance to the local community. In the last

two decades, date palm cultivation in the UAE has experienced remarkable growth, with over 250 varieties of the crop being farmed in various parts of the nation, especially in Al Ain and Abu Dhabi emirates. The total growth rate of date fruit is paralleled by a high consumption rate in the UAE, with a per capita daily intake of 10 to 200 g in Abu Dhabi emirate alone (Habib & Ibrahim, 2011). Khesab cv. can be consumed at Rutab and tamer stage. Khesab cv. has a slow ripening which means that it has late maturity (Ahmed et al., 2021a). Therefore, slowing down the ripening process and retarding fruit senescence is critical for enhancing the marketability and storage life when it is not the season where it is consumed. To the best of our knowledge, there are no reports on the synergistic effects of different natural elicitors on Khesab cv. fruit particularly as preharvest spray treatments.

1.6 Research Objectives

- (a) To explore the synergistic effects of natural elicitors such as chitosan, calcium chloride and salicylic acid as preharvest spray treatments on fruit quality and marketability of Khesab cv. variety fruits.
- (b) Study of the effect of chitosan in combination with calcium chloride and/or salicylic acid on the shelf life and physicochemical constituents of Khesab cv. fruit.
- (c) Investigation of the effects of chitosan in combination with calcium chloride and/or salicylic acid on the storability of fruit of the variety Khesa during two months of cold storage.

Chapter 2: Literature Review

2.1 Introduction

In general, date palm and all crops can get affected by diseases that can affect the fruits and the shelf life of the crops. The use of chemical pesticides and treatments helps preserve the quality of the fruit and extend shelf life. However, this method has been facing some challenges due to the phytotoxicity of this chemical preservative, development of antimicrobial resistance, and food safety concerns regarding the effects of chemical residues on human health and the environment (Shen & Yang, 2017). Therefore, safer, more effective, economical, and convenient storage methods are needed to continue production without affecting human health and the environment (Hassani et al., 2004). The main aim is to extend the shelf life of the fruit, and one of the methods to achieve this goal is using natural elicitors (Baldwin et al., 2011). Elicitors can mimic the action of the signalling molecules, activate defence responses and trigger and mediate long-lasting systemic acquired resistance (SAR), which protects against a wide range of pathogens that causes fruit decay (Baenas, 2014; Chen et al., 2018; Moreno-Escamilla et al., 2018).

2.2 Chitosan as Natural Elicitors

In recent decades, the biodegradability of natural compounds from animals and plants has attracted the interest of plant pathologists. Many studies have been conducted to delay the postharvest quality changes of date fruits. An alternative approach is to use natural triggers to enhance physiological adaptations and accelerate the plant defence system rather than different approaches such as cold storage, modified atmosphere packaging (Alsawmahi et al., 2018), and controlled atmosphere storage (CA) (Baloch et at., 2006; Baenas et al., 2014; Alhamdan et al., 2015; Aleid & Saikhan, 2017; Alsawmahi et al., 2018; Moreno-Escamilla et al., 2018). Chitosan has received increasing attention due to its fungicidal action and triggering of defence mechanisms in plant tissues (Terry & Joyce, 2004). Chitosan is an N-acetylated derivative of the polysaccharide chitin. It is a natural polymer with a polycationic nature. Chitosan has numerous agricultural applications (e.g., soil conditioner, film, fungicide, trigger), agroindustry, cosmetics, environmental protection, biomedicine, and wastewater management) (Katiyar et al., 2014). Many studies have shown the high potential of chitosan for preserving fresh fruits and vegetables (Rhoades & Roller, 2000).

2.2.1 Preharvest application of chitosan

Several studies on pre-and postharvest applications have shown proven success with various fruit crops using environmentally friendly triggers, such as chitosan (e.g., Romanazzi et al., 2013; Shen & Yang, 2017; Romanazzi et al., 2018; Rahman et al., 2018; Abu-Shama et al., 2020), and salicylic acid (Zhang et al., 2003; Sayyari et al., 2009; Luo et al., 2011; Mohamed et al., 2014; Chen et al., 2020; Gomes et al., 2021), and calcium chloride (Irfan et al., 2013; Sohail et al., 2015; Atia et al., 2018). These elicitors lead to retarding tissue softening, delaying ripening, enhancing quality, extending shelf life, and preventing fruit deterioration. In addition, pre-harvest spraying with chitosan is highly functional and has a beneficial effect on fruit quality attributes (Reddy et al., 2000).

The effect of chitosan on controlling plant diseases and pathogenic microorganisms stated that for achieving the goal of sustainable agriculture, chitosan would become a popular plant protectant (Hassan & Chang, 2017). The effects of natural elicitors on the quality of lettuce showed that the application of chitosan exerted a fungistatic effect

that reduced the yeast and mould population throughout storage and decreased the activities of PPO and POD, enzymes related to browning processes. Microscopical observations indicate that chitosan directly affects the morphology of the chitosan-treated microorganism, reflecting its fungistatic or fungicidal potential. Other studies strongly suggest that chitosan induces a series of defence reactions correlated with enzymatic activities (e.g., Banos et al., 2006).

Preharvest application of chitosan onto kiwi fruit has decreased the weight loss and delayed the changes in physicochemical properties that include firmness, soluble solids content, titratable acidity, total sugars, total acids, total phenols, and total lignin (Kumarihami et al., 2021). The preharvest foliar application of 3% chitosan with a postharvest coating of aloe vera gel extended the storage life of table grapes up to 15 days by reducing decay index, malondialdehyde content, weight loss and polyphenol oxidase (Nia et al., 2021). Pre-harvest application of chitosan is beneficial for improving strawberries' quality and antioxidant capacity (He et al., 2018). The treatment of chitosan in combination with calcium chloride (Ca-chitosan) at the preharvest stage in kiwi fruit resulted in enhancing fruit quality and postharvest life (Kim, 2018).

Shen and Yang (2017) developed chitosan-g-salicylic acid (CTS-g-SA) for preharvest treatment of grapes; CTS-g-SA treatment exhibited enhanced activities of phenylalanine ammonia-lyase, chitinase, and β -1, 3-glucanase, while also promoting the accumulation of phenolic compounds and more excellent resistance to Botrytis cinerea decay. The preharvest treatment of strawberries with chitosan, laminarin, extracts of Abies spp., Polygonum spp., Saccharomyces spp., calcium and benzothiadiazole provided ~ a 30% reduction in postharvest decay against grey mould

and Rhizopus rot, without affecting fruit colour and firmness (Feliziani et al., 2015). Previous studies have shown that chitosan reduces decay incidence, mainly caused by Botrytis cinerea in tomatoes (El Ghaouth et al., 1992), decreases microbial count, and elicits (Kibar & Sabir, 2018). In another experiment, chitosan significantly reduced the disease lesion size on tomato fruit when the tomato plants were pre-treated with 1.0% or 2.5% (w/v) COS solution ten days before being inoculated with Collectorrichum sp. (Munoz et al., 2009).

2.2.2 Postharvest applications of chitosan and their derivatives

Increasing polyphenol oxidase and peroxidase enzyme activities, change in acidity, anthocyanin and minimising the growth of mesophilic aerobic yeasts. Moulds populations can be achieved when blueberries are coated with chitosan, silicon and titanium dioxides (Li et al., 2021). Treating mangos by coating them with chitosan in combination with aloe vera gel leads to suppressed diseases, maintains mango fruit's natural properties during postharvest storage, and extends the storage life of mango fruit (Shah & Hashmi, 2020).

Chitosan composite film (CCF) containing chitosan, dextrin, ferulic acid and calcium effectively reduced the incidence of soft rot in kiwi fruit caused by fungi *B. dothidea* and Phomopsis sp. In addition, the CCF film increased the content of resistance compounds, and the activity of defence enzymes enhanced the yield and quality and prolonged the shelf life of kiwifruit (Zhang et al., 2020). Furthermore, Khatri et al. (2020) indicated that the efficiency of aloe vera gel and chitosan coatings in prolonging the postharvest- the life of tomatoes showed that the composite coating of chitosan and aloe vera gel treatment delayed the ripening process and extended the shelf life of tomatoes up to 42 days (Khatri et al., 2020).

Nguyen and Nguyen (2021) reported the influence of calcium chloride with nano chitosan coating on the quality of strawberries during postharvest storage resulted that chitosan coating with calcium chloride reduced weight loss, maintained L-ascorbic acid, total anthocyanin, antioxidants, retarded malondialdehyde production and no bitterness detected in the treated strawberries on 15 d of storage at 4°C. Studies reported that chitosan could be more effective in the postharvest physiology of fruit crops when it is combined with other additives, including salicylic acid (Huang et al., 2017), potassium silicate (Mohamed et al., 2017), calcium chloride (Kim, 2018), gum Arabic (Khaliq et al., 2016), lactoperoxidase (Cissé et al., 2015), and antagonistic yeast (Meng & Tian, 2009). After harvesting coating with 1.0%, chitosan showed that the quality attributes of apple fruits could be maintained for up to 80 days during storage at $18\pm2^{\circ}$ C and $56\pm2\%$ RH (Zeb et al., 2020).

Saki et al. (2019) reported that fig fruit coated with chitosan and thymol essential oil had less fungal decay incidence, lower weight loss and respiration rate. In addition, the coated fruit exhibited shelf-life extension with higher firmness, total soluble solids and anthocyanin content after twenty days of storage at 6°C. A nanocomposite coating of chitosan and MMT (a natural clay) has been developed and applied to tangerine fruits (Xu et al., 2018). The results showed that adding 1% (w/w) MMT reduced the water sensitivity and significantly improved the oxygen barrier properties of chitosan films, thus providing longer postharvest life for tangerine fruits (Xu et al., 2018).

Using chitosan and calcium chloride individually and in combination as a pretreatment of guava fruit leads to a significant delay decline in physiological loss of weight, enhanced total soluble solids firmness, acidity, and ascorbic acid, sugars, phenols and total antioxidant activity during storage (Chawla et al., 2018). In addition, chitosan, potassium silicate, and calcium chloride coating on mangos showed an increase in fruit storability and shelf life compared to the mangos treated individually or not been coated from the beginning (Mohamed et al., 2017).

Romanazzi et al. (2018) reported that using the treatment of chitosan mixed with ethanol, wax, and similar types of organic materials improved the protecting effect on grapes from grey mould compared to the application of using chitosan alone. Furthermore, Gayed et al. (2017) reported that 2% CaCl2+1% chitosan was most effective in minimising weight loss (%) and decay (%), as well as in maintaining maximum firmness and lengthening the shelf life of peach fruits. Furthermore, when using chitosan integrated with plant derivatives like plant extracts and essential oils, organic salts and acids, and antagonistic microorganisms, including yeast and bacteria, results show that it effectively reduces postharvest fungal rots (Bautista-Baños et al., 2017).

Strawberry fruits coated with 1% and 2% chitosan solution and stored at 2°C for nine days showed reduced water loss and delayed the qualitative changes in colour, titratable acidity and ascorbic acid content. (Petriccione et al., 2015). Using chitosan as a postharvest treatment has been studied for efficacy in inhibiting decay and extending the shelf life of perishable produces such as plum (Kumar et al., 2017), peach (Elbarbary & Mostafa, 2014), fresh-cut melon (Poverenov et al., 2014), strawberry (Wang & Gao, 2013) and cucumber (Ben-Shalom et al., 2003). The use of chitosan integrated with essential oil including lime, thyme, bergamot, clove and cinnamon leads to a reduction of the appearance of fungus *B. cinerea, C. gloeosporioides, P. digitatum* and *Phytophthora drehsleri*, and yeasts and moulds on

strawberries, figs, avocado, cucumber, mandarin and grapes (Mohammadi et al., 2015).

Coatings, the combination of chitosan with oleic and acetic acid, reduced disease symptoms for ten days. They decreased decay caused by fungi and bacteria in strawberries and prickly pears (Ochoa-Velasco & Guerrero-Beltrán, 2014, Xylia et al., 2021). Velickova et al. (2013) reported that chitosan-beeswax coatings decreased the senescence and weight loss of strawberries, modified the respiration rates and slowed down metabolism by retaining the colour and the texture of the fruit. Combining 1% chitosan with 3% ammonium carbonate solutions and 2.5% calcium reduces the severity of anthracnose and incidence in papaya during 14 days of storage (Al-Eryani-Raqeeb et al., 2009). Coating strawberry 'Camarosa' with 1.5% chitosan and 0.5% calcium gluconate showed no visible symptoms of disease caused by moulds during the whole storage period at 10°C (Hernández-Muñoz et al., 2008).

2.2.3 Treatments with chitosan and combination of calcium chloride and salicylic acid

Chitosan and salicylic acid treatment on fresh pistachio enhanced fruit quality in refrigerated storage with prolonged shelf life. The fruit was lighter, redder, and more yellow with the highest sensory scores for colour, texture, and overall acceptance with reduced growth of bacteria and fungi (Molamohammadi et al., 2020, Ahmed et al., 2021b). Preharvest treatments of calcium spray with the postharvest treatments of chitosan coating on the Chinese dwarf cherry resulted in an extension in the storage time and reduced fruit decay rate at the end of storage. In addition, they improved the quality of the fruit for up to 30 days (Guo et al., 2020). Treating the fresh-cut apples

with chitosan combined with calcium chloride showed lower weight loss, improved appearance reduced browning and enhanced ascorbic acid content (Liu et al., 2016).

The integrity of soluble pectin is shown to be maintained in the fresh-cut honeydew melon when using the edible coating of chitosan combined with calcium chloride that extends the shelf-life of the fruit (Ahmed and Palta 2011, Chong et al., 2015). Furthermore, postharvest treatment of pomegranate with salicylic chitosan exhibited lower weight loss, respiration rate and ethylene production associated with higher firmness, total soluble solids, and titrable acidity as sensory quality (Sayyari et al., 2016). In addition, Vyas et al. (2016) reported that salicylic acid treatment combined with calcium chloride and sodium benzoate effectively enhanced antioxidants, ascorbic acid, and total anthocyanins in phalsa fruit (*Grewia asiatica* L.). More treatments of chitosan in combination with other treatments is reviewed in Table 1.

Fruit	Physiological change	Combination	Reference
		with chitosan	
Apple	• Total phenolic, flavonoids,	Olive waste	Khalifa et al.,
	antioxidants, pigments, weight	extract	2017
	loss		
Peach	Malondialdehyde content	Gamma-ray	Elbarbary &
	• Total soluble solids, weight	Silver and zinc	Mostafa,
	loss, ascorbic acid content	oxide	2014.
	• Colour and fruit firmness	nanoparticles	Kaur et al.,
		Polyethylene	2017;
		terephthalate	Cindi et al.,
		punnets	2015
		containing	
		thyme oil	
Plum	• Fruit firmness, weight loss,	Calcium	Gayed et al.,
	total soluble solids, total	chloride	2017
	phenolic content, and		
	titratable acidity		
	• Respiration rate, fruit colour,	Ascorbic acid	Liu et al.,
	polygalacturonase, superoxide		2014
	dismutase, peroxidase,		
	catalase, polyphenol oxidase		

Table 1: Physiological changes in fresh fruits after chitosan treatment and with other elicitors

Fruit	Physiological change	Combination	Reference
		with chitosan	
Sweet	Malondialdehyde	-	Pasquariello et
cherry	content and antioxidant		al., 2015
	enzymes		
Citrus	• Fruit firmness, weight	Carboxymethyl	Arnon et al.,
	loss, total soluble solids	cellulose	2014
	• Peroxidase and	Cyclic	
	phenylalanine	lipopeptide	Waewthongrak
	ammonia-lyase	antibiotics from	et al., 2015
		Bacillus subtilis	
Mango	• Peroxidase (POD) and		Gutierrez-
	polyphenol oxidase		Martinez et al.,
	(PPO) gene expression		2017
Kiwi	Induced gene		Zheng et al.,
fruit	expression and		2017
	increased enzymatic		
	activity of catalase,		
	superoxide dismutase		
	and ascorbate		
	peroxidase		

Table 1: Physiological changes in fresh fruits after chitosan treatment and with other elicitors (Continued)

Fruit	Physiological change	Combination with	Reference
		chitosan	
Strawberry	• Weight loss, titratable	Carboxymethyl	Gol et al., 2013
	acidity, pH, total soluble	cellulose	Sangsuwan et
	solids, total phenols	Lavander and	al., 2016
	• Weight loss	thyme essential oil	
	• pH and soluble solids	Natamycin, nisin,	Duran et al.,
	content	pomegranate, grape	2016
	• Weight losses, total	seed extract	
	soluble solids and	Olive waste extract	Khalifa et al.,
	titratable acidity		2016
Grapes	Phenylalanine ammonia-	Salicylic acid	Shen and Yang,
	lyase, chitinase, and β -1,		2017
	3-glucanase, phenolic		
	compounds		Al-Qurashi and
	• Total phenols,	Menta essential oil	Mohamed, 2015
	flavonoids and ascorbic		Guerra et al.,
	acid content, activities of		2016
	peroxidase		
	• Firmness, titratable		
	acidity, soluble solids,		
	colour, weight loss		

Table 1: Physiological changes in fresh fruits after chitosan treatment and with other elicitors (Continued)

Fruit	Physiological change	Combination with	Reference	
		chitosan		
Pear	• Total phenolic and	Calcium chloride	Kou et al.,	
	flavonoid contents,		2014a	
	superoxide dismutase,			
	peroxidase and catalase	Calcium chloride		
	activities, total		Kou et al.,	
	antioxidant activity		2014b	
	• Malic acid-metabolising			
	enzymes and related			
	genes expression			

Table 1: Physiological changes in fresh fruits after chitosan treatment and with other elicitors (Continued)

(Source. Romanazzi., et al. 2018)

According to Lin et al. (2021), the various stages describe the cell division, juvenile green, the experienced hard full shaded, the delicate brown, and the firm raisin-like organic product. As stated by Khesab cv. It is an exceptionally regular cultivated product in the region of the UAE, and it is consumed at the Rutab stage, which is half the delicate stage of the product. After collecting, the natural product begins losing its solidness, shading, taste, and generally speaking physiology, which finally brings down the commonly estimated quality of the fruits. These changes are results of the post-harvest action of different ageing-related chemicals and chemicals that mellow the natural product tissue, decline the immovability and genuine uprightness of the organic product, and also, at last, lead to the natural product's senescence.

According to Hinkaew (Hinkaew et al., 2020), salicylic acid is considered a phenolic factor that increases the ability to prevent disease and postpones the ripening stage of the fruit. At the same time, calcium is also considered an essential element that delays the maturity of the fruits. Therefore, the application of these harmless chemicals can increase the longevity of the fruit, which is the pre- and post-harvesting scenario of the dates. According to Kumarihami et al. (2020), during the pre-cultivating stage pre- and post-harvesting stages, the application of the solution (Ch) 1% with salicylic acid (SA) and calcium chloride (Ca) (the solution can be in the combination of Ch, SA, Ca, Ch+Ca, Ch+SA, Ch+SA+Ca) can increase the quality of the products. According to Elmenofy (2021), natural and eco-friendly components like chitosan, calcium chloride, and salicylic solution can increase the tissue softening of the dates, visibly delaying the ripening stage and restoring quality, extending the shelf life of the dates and preserving them properly.

According to Badawy (2019). as chitosan, calcium chloride is considered a natural solution that strengthens the tissues of the trees and fruits. It is a natural solution applied in pre- and post-harvest situations to Khesab cv. Dates and increase its prospects of taste and quality. At the same time, calcium is considered the element of tissue softening and an active agent that delays the ripening stage of the Khesab cv. dates. Abdel-Hamid (2020) states that the solution of chitosan, calcium chloride, and salicylic solution can increase the quality of the Khesab cv. dates which tend to lose their taste and quality if it is ripened enough. It is essential to increase the self-life and market life of the fruits to increase their economic value. These solutions can provide natural and eco-friendly support to plant-based harvesting and increase the self-life and quality of the fruits. Thus, in the pre-and post-harvesting situation, the farmers implement these solutions to incorporate various benefits and sustain the quality of the dates to increase their market prospects.

Chapter 3: Materials and methods

3.1 Plant Material and Sampling

During the 2020 season, uniform date palm trees (*Phoenix dactylifera* L., cv. Khesab) were randomly selected in the experimental farm of the College of Food and Agriculture located in Al Foah region, Al Ain, UAE, located in the co-ordinate latitude and longitude of 24.2191°N and 55.7146°E. In addition, there were eight female spathes per palm tree, and the trees were pruned for an 8:1 leaf to bunch ratio. The design of the experiment was a randomized complete block design with six palms (replicate) from each 'Khasab' palm receiving seven different treatments (one treatment for each bunch).

Three stages of development (5 and 15 weeks from pollination and two weeks before harvest) were selected for spraying treatments with different elicitors: chitosan, SA, and CaCl2 alone and in combinations (7 treatments): Control (Water), Ch, SA, Ca, Ch+Ca, Ch+SA, Ch+SA+Ca with adding 0.2 of between 20 which help in reducing the tension of the water and create a soap texture that helps the treatments in holding onto the date fruit. The control date palm trees were sprayed with only deionized water for control. The harvesting of bunches was done at the commercial stage (when roughly about 50% of dates have reached the Rutab stage (half Bisr).

After washing, the fruit was carefully separated according to their maturity level (Bisr and 50% Rutab (so-called Rutab). Then Bisr fruit was used for further analysis. Randomly, 100 fruit were collected from each treatment for initial physicochemical, phytochemical, bioactive properties, and microbial analysis at harvest time (day 0) (Awad & Al-Qurashi, 2012). Next, another batch of 500 fruit from each treatment was collected and stored in punched plastic bags (100 fruit in each bag) and stored at 2°C and 90–92% relative humidity (RH) for 60 days. For 15 days, a bag was withdrawn randomly for analysis for each time interval. According to Hinkaew (Hinkaew et al., 2020), salicylic acid is considered a phenolic., as shown in Table 2.

Treatment	Chemicals	Application	
Ch	Chitosan	1%	
Ca	Calcium chloride	3%	
SA	Salicylic acid	2 mM	
Ch+SA	Chitosan+Salicylic acid	1:1, <i>v/v</i>	
Ch+Ca	Chitosan+Calcium chloride	1:1, v/v	
Ch+Ca+SA	Chitosan+Calcium chloride+Salicylic acid	1:1:1, <i>v/v/v</i>	

Table 2: Preharvest spry treatments of Khesab cv. date fruit with different elicitors

3.2 Physiochemical analysis

3.2.1 Fruit characteristics

Fruit dimension studies were conducted after harvest according to Raste-gar et al. (2012). Fruit weight loss during storage was recorded every two weeks for each treatment and expressed as percent weight loss from the original weight before cold storage, using the following equation:

3.2.2 Fruit ripening and decay percentage

A visual evaluation of fruit was done to record the ripening burst (fully *Rutab*) and spoilage every other week until a complete decay of fruit occurred during storage for 60 days. The ripening and decay percentages were calculated using the following equation (Kumar et al., 2013):

Decay or ripe fruit (%) = <u>Number of ripe or number of decay fruit</u> x 100

Total fruit number

3.2.3 Total soluble solids (TSS)

Ten grams of pitted date fruit were pureed and mixed with 10 mL of distilled water in a blender to form a slurry, which was filtered to obtain the clear juice. The Brix value was determined in the juice using a digital refractometer (DR 6000, A. Kruss Optronic GmbH, Hamburg, Germany).

3.2.4 Fruit surface colour

The surface colour of fruits was determined by using a Hunter Lab colourimeter (Hunter Lab Inc., Reston, VA, USA).

The values were represented as L* (brightness), a* (blue/yellow), and b* (red/green) (Ferrer et al., 2005). These values were further used to calculate the Hue angle ((h°), chroma (C*), and total color difference (ΔE^*) as follows; ho = 180° + arctan (b*/a*); (C*) = (a*2 + b*2)1/2.

 $\Delta E = [(L^*-L^*0) + (a^*-a^*0) + (b^*-b^*0)]1/2 \text{ (Maskan, 2001), where } L^*0, a^*0, and b^*0$ values were from control fruit at harvest time (day zero) (Feliziani et al., 2015).

3.2.5 Determination of total microbial load

The total fungal/mould and bacterial count on fruit samples were determined upon harvest (day 0) and at the end of storage (60 days). One gram of mashed fruit tissue was vortexed vigorously in 9 mL autoclaved buffered peptone water under sterile conditions and serially diluted to prepare several different dilutions. Employing the pour plating technique, a 1 mL sample from each dilution was plated with plate count agar (PCA) which was incubated at 37°C for 48 h and potato dextrose agar (PDA) incubated at 27°C for 5 d. The number of colonies was recorded at the end of incubation time and calculations were performed using Log10 colony-forming unit per g of fresh weight (Log10 CFU g⁻¹) (Xylia et al., 2021)

3.2.6 Extraction of bioactive compounds

The extraction of phenolic compounds was performed via homogenizing two grams of fresh fruit samples in 20 mL of 80% methanol with a water bath shaker at 150 rpm for 24 h, 45°C. Whatman no.1 filter paper was used to filter the slurry. The filtrate (date extract) was used for further analysis (Ahmed et al., 2021a).

3.3 Phytochemical Analysis

3.3.1 Total Phenolic Content (TPC)

Following (Velioglu et al., 1998, Ahmed et al., 2021c), total phenolic content was estimated with some modifications. First, date extract (100 μ l each) was added to clean and dry test tubes, followed by 50 μ l Folin Ciocalteu reagent, and then vortexed. Then,

all tubes were incubated at room temperature for 2 min. Then, 2 mL of NaOH (6%) was added to each tube and incubated in the dark for 45 minutes. Finally, absorbance was measured at 750 nm using a UV spectrophotometer. Results were expressed as milligrams of gallic acid equivalents (GAE) per 100 g fresh weight (mg 100 g-1 GAE) according to the standard curve obtained by measuring the absorbance of known gallic acid concentrations.

3.3.2 Total Tannin Content (TTC)

The amount of tannin in the date extracts was determined using the colourimetric method described by Bentebba et al. (2020), with some modifications. First, 1 mL of a 4% vanillin solution prepared in absolute ethanol and 0.2 mL of HCl (37%) was added to 0.4 mL of extract or catechin as a standard. The mixture was then shaken and allowed to react in the dark at room temperature for 15 min before absorbance was measured at 500 nm using a spectrophotometer. Total tannin content was expressed as milligrams of catechin equivalents per 100 g fresh weight (mg 100 g-1 CE) (Ahmed et al., 2021c).

3.3.3 Total Flavonoids Content (TFC)

According to Kim et al. (Kim et al., 2003), total flavonoids were determined with some modifications. First, 75 μ L NaNO2 (5%) was added to the data extract (250 μ L) in a test tube and vortexed, followed by 5 min incubation in the dark. Next, 75 μ L of AlCl3 (10%) was added; the mixture was vortexed and kept in the dark for 6 min, followed by 500 μ L of NaOH (1 M), then the volume was made up to 2.5 mL with distilled water. Then, the absorbance was measured using a spectrophotometer (Shimadzu,

Kyoto, Japan) at a wavelength of 510 nm. Results were expressed as milligrams of catechin per 100 g fresh weight (mg 100 g-1 CE).

3.4 Antioxidant Activities

The evaluation of the antioxidant activities was based on the method developed by Abd Elwahab et al. (2019).

3.4.1 DPPH radical scavenging activity

DPPH radical scavenging activity was determined according to Ahmed et al. (2022b) method with slight modifications. The 1.5 mL samples with concentrations ranging from 0.5 mg to 10 mg L-1 were mixed with 1.5 mL of 0.15 mM 2,2-diphenyl-1-picrylhydrazyl (DPPH) in 95% ethanol. After incubation for 30 min at room temperature in the dark, the values were measured at 517 nm using a spectrophotometer. The blank sample was prepared similarly for each concentration, except that ethanol was used instead of DPPH solution. IC50 values (mg mL-1) were calculated for each sample.

3.4.2 ABTS radical scavenging activity

The activity of the ABTS radical scavenger was determined as follows. Stock solutions of ABTS (7.4 mM) and potassium persulfate (2.6 mM) were prepared and stored in a refrigerator (4°C). Equal parts of the two stocks were mixed and allowed to react for 12 hours at room temperature in the dark. Before performing the assay, the solution was further diluted by mixing 1 mL of the ABTS solution with 50 mL of methanol, which gave an absorbance of 1.1 ± 0.02 units at 734 nm, as measured by a spectrophotometer. A new ABTS solution was prepared for each assay.

Then, 150 μ L of the sample with concentrations ranging from 0.5 mg to 10 mg mL-1 was mixed with 2850 μ L of ABTS solution and left in the dark at room temperature for two hours. The absorbance was then measured at 734 nm using a UV spectrophotometer. The blank was prepared in the same manner for each concentration, except that methanol was used instead of ABTS solution. A Trolox standard curve was prepared in the range of 50 μ M to 600 μ M. Activity was expressed as milligrams of Trolox equivalents (TE) per 100 g fresh weight (mg 100g-1 TE).

3.5 Statistical Analysis

Analysis of variance (one-way ANOVA) was employed for the completely randomized design experiment with six replicates using SAS statistical software (SAS Institute Inc., 2000, Cary, NC, USA). Least significant differences (LSD) at level $p \le$ 0.05 were utilized to compare means between treatments at each time interval for each analysis. The correlation coefficient between the main biochemical and physical characteristics was calculated by SAS.

Chapter 4: Results and Discussion

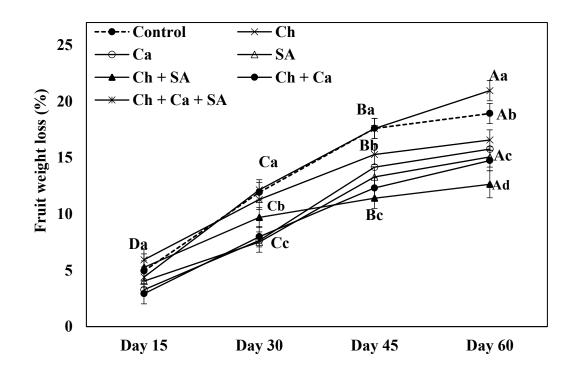
4.1 Physical Quality Characteristics of Fruit at Harvest

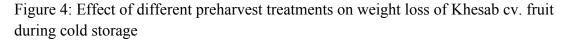
The average weight, width and length of fruit of cultivar Khesab at harvest are shown in Table 3 - significant differences (p < 0.05) in physical characteristics of treated cultivar Khesab. Fruit was affected by the preharvest elicitor treatments: the average weight, width and length of Khesab cv. Fruit was significantly (p < 0.05) higher in the SA and Ch+ SA treatments than in the control and the other treatments. These improvements in fruit physical characteristics could be due to the influence of SA in combination with Chin, which maintains cell integrity and increases firmness and improves fruit weight and size (Kassem et al., 2010). Similarly, Mohamed et al. (2014) described that preharvest spray application of SA significantly increased the weight, width, and length of date fruits compared to control fruits. The physical characteristics of the fresh fruit are of critical importance as they directly indicate the physical effects of the applied elicitors. The above results show that the application of SA alone or in combination with Ch affected the physical properties of fresh fruits of Khesab cultivar.

4.2 Fruit Weight Loss during Storage

Weight loss is a significant factor that reduces postharvest fruit shelf life and marketability (Kassem et al., 2010; Mohamed et al., 2014). The results showed that the fruit weight of cultivar Khesab was significantly (p < 0.05) affected by the application of triggers. All fruits experienced continuous weight loss during storage, which varied among treatments. However, significant differences in weight loss were evident between the treatments and the control. After 60 days of storage, the least weight loss was observed in fruits treated with Ch+ SA, followed by SA, Ch+Ca, Ca,

Ch with 12.64, 14.74, 15.06, 15.76, 16.57%, respectively. The weight loss during storage may be due to an increase in respiration rate and/or overall moisture loss from the fruit (Shiri et al., 2013; Mohamed et al., 2014; Atia et al., 2018). Figure 4 shows the per cent weight loss that occurred during cold storage at 20°C and 90-92% RH for 60 days in the harvested fruits.





Note: Values are the mean $(n = 25) \pm SE$. Means with different letters between treatments (small letters) at a different time interval (capital letters) are significantly different at p < 0.05using the LSD test. Ch: chitosan; Ca: calcium chloride; SA: salicylic acid.

Table 3 shows the potential effects of different preharvest treatments on Khesab cv. fruit physical characteristics at harvest.

	Fruit Weight	Fruit Length	Fruit Width
	(g)	(mm)	(mm)
Control	$10.2 \pm 0.11b$	$35.0 \pm 0.38b$	$23.5 \pm 0.24b$
Ch	$10.9 \pm 0.22b$	$35.2 \pm 0.36b$	$24.1\pm0.27ab$
Ca	$10.0 \pm 0.22b$	$34.7\pm0.38b$	$23.9 \pm 0.23b$
SA	$11.8 \pm 0.25a$	$36.7 \pm 0.40a$	$24.2 \pm 0.25 ab$
Ch + SA	$12.4 \pm 0.46a$	37.2 ± 0.48 a	$25.6 \pm 0.32a$
Ch + Ca	$10.8\pm0.39b$	34.9 ± 0.51 b	$24.2\pm0.34ab$
Ch + Ca + SA	$10.6 \pm 0.16b$	$35.2 \pm 0.33b$	$24.2\pm0.21ab$

Table 3: potential effects of different preharvest treatments on Khesab cv.

Note: Values are the mean (n = 25) \pm SE. Means with different letters between treatments (small letters) at a different time interval (capital letters) are significantly different at *p* < 0.05 using the LSD test. Ch: chitosan; Ca: calcium chloride; SA: salicylic acid.

The low weight reduction observed in fruit sprayed with chitosan alone might be due to the film-forming properties which significantly decreased the evaporation rate of water from the fruit, as noted in the case of other fruit treated with chitosan (Petriccione et al., 2015; Petriccione et al., 2015; Romanazzi et al., 2018, Maan et al., 2021). Also, Ca and SA were found to have significantly lower weight loss compared to control in constant to Atia et al. (Atia et al., 2020) reported that pre-storage treatment with Ca and SA reduced weight loss in 'Barhi' dates at *Khalal* stage. Also, Kassem et al. (2010)

and Mohamed et al. (2014) found that the preharvest application of SA reduced weight loss during storage in treated Khesab cv. fruit compared to the control. SA is well known to reduce chilling injuries, inhibit ripening, and act against various abiotic and biotic factors (Ennab et al., 2020).

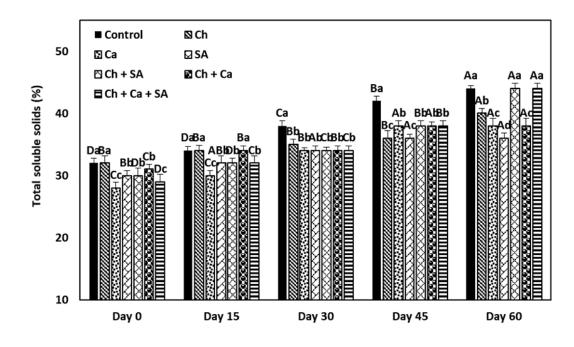
This suggests the positive physiological condition of fruit treated with SA presumably owing to decreased respiration rate that can also be associated with the improved fruit turgidity. In dates, typically, the weight of the fruit increases as the maturation advance and at its peak at the *Khalal* stage, then promptly decreases as ripening progresses (Alqarni & Bazzi, 2019). Based on the above results, the weight loss reduction observed in fruit treated with Ch alone or in combinations with SA and Ca, indicates the good physiological conditions of the treated fruit, most likely as a result of the reduced respiration and transpiration rates, and the regulating effects of these elicitors on the ripening process.

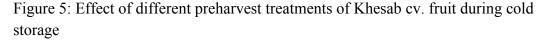
4.3 Total Soluble Solids (TSS) during cold storage

Overall, the concentration of TSS gradually increased in all fruits as ripening progressed during cold storage. At harvest and after 60 days of cold storage, control fruits had the highest TSS concentration (from 32 to 42%), followed by Ch+ SA treated fruits (Feliziani et al., 2015). Ca treated fruits exhibited the lowest TSS concentration (from 28% to 38%). Similarly, Ch+Ca and SA treated fruits had significantly (p < 0.05) lower levels of TSS compared to control fruits after 15, 30, 45 and 60 days of cold storage (Figure 5). It has been reported that elicitor efficacy is determined by fruit tissue reactivity, which decreases as ripening progresses (Romanazzi et al., 2018). In this study, the observed TSS concentrations were similar to those described by Kassem et al. (2010) in SA treated date fruit. However, Mohamed et al. (2014) reported no

significant difference between SA treated date fruits and the control fruits in terms of TSS content.

The results of Ca treatment might be due to the high Ca concentration (3%) in the treatment. The usual increase in TSS during cold storage could be due to the degradation of large polysaccharide molecules into smaller sugars by enzymatic activities and water loss (Mohamed et al., 2014; Hazbavi et al., 2015). The TSS concentrations showed significant differences (p < 0.05) between the different treatments, as shown in Figure 5. These results are consistent with the weight loss results shown in Figure 4. According to our findings, the degradation of polysaccharides was slowed down by Ca alone or in combination with Ch and SA, resulting in lower TSS concentrations in the treated fruits and, consequently, in a lower ripening rate.





Note: values are the mean (n = 25) \pm SE. Means with different letters between treatments (small letters) at a different time interval (capital letters) are significantly different at p < 0.05 using the LSD test. Ch: chitosan; Ca: calcium chloride; SA: salicylic acid.

4.4 Fruit Ripening

The results of fruit ripening during cold storage at 2°C for 60 days are shown in Figure 6. The different treatments significantly delayed fruit ripening during the storage period (p < 0.05). In general, the rutab content gradually increased during cold storage in all treatments. At the end of storage period, the highest rutab content was observed in control fruits (90.3%), compared to SA, Ch+ SA, Ch+Ca, Ch, Ch+ SA +Ca and Ca treated fruits with a range of 79-83%. Ch+ SA +Ca treatment alone significantly delayed normal ripening of Bisr fruit compared to the other treatments in combination with Ca. These results are consistent with those of TSS in Figure 5, where fruit treated

with Ca, Ch+Ca, Ch+ SA +Ca, and SA had lower TSS content than the control and the other treatments. Moreover, fruit maturity (Rutab %) was positively correlated with the increase in TSS and weight loss (Table 4).

These observations may also indicate that the elicitors delay the ripening of Khesab cv. fruit during cold storage. It is well known that calcium administration slows down the respiration rate and delays fruit ripening (Irfan et al., 2013; Sohail et al., 2015; Atia et al., 2018). Similarly, preharvest application of SA delayed date fruit ripening by three weeks compared to control fruits (Kassem et al., 2010). In addition, pre- and postharvest treatments with SA were reported to improve the quality and shelf life of many fruits and prevent their spoilage (Pila et al., 2010; Shen & Yang, 2017; Atia et al., 2018). In addition, fruit ripening can be slowed by the film-forming ability of Ch, creating a barrier to gas exchange and reduced respiration. It should be noted that less mature fruits are less susceptible to postharvest rot (Romanazzi et al., 2018). Figure 6 illustrates various preharvest treatments on Khesab cv. In ripening stage "*Rutab*".

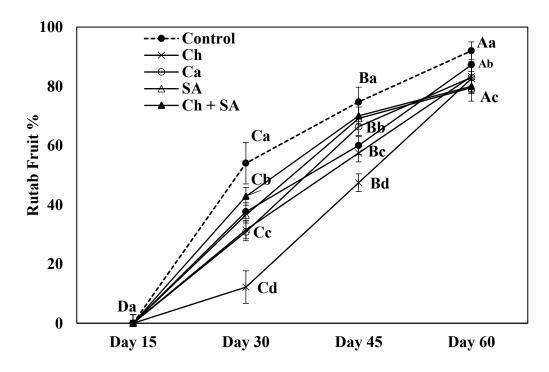


Figure 6: Effect of different preharvest treatments on Khesab cv. fruit ripening during cold storage

Note: Values are the mean $(n = 25) \pm SE$. Means with different letters between treatments (small letters) at a different time interval (capital letters) are significantly different at p < 0.05using the LSD test. Ch: chitosan; Ca: calcium chloride; SA: salicylic acid.

Fruit ripening is a natural phenomenon that leads to membrane impermeability and gradual senescence of fruit tissue due to oxidation of essential membrane components (Awad et al., 2011b). Ethylene is the main hormone responsible for the processes of ripening and senescence (Kassem et al., 2010). It is possible that the sprayed elicitors can control the production or action of ethylene, thereby slowing down fruit ripening (delaying the onset of rutab). Our results showed the retarding effect of Ch, Ca and SA

in different combinations on fruit ripening compared to the control. Nevertheless, date ripening is not an asynchronous process for all date fruits in the same cluster, as there may be fruits at different stages of ripening in the same cluster at any given time. Bisr fruit that received the same treatment could enter the rutab stage later than control fruit in the same cluster. As a result, different significant biological variations between individual fruits within the same cluster may be responsible for certain contradictory results (Mohamed et al., 2014).

4.5 Fruit Decay During Storage

The results of the percentage of fruit deterioration during cold storage at 2°C for 60 days are shown in Figure 4. It was found that the different preharvest spray treatments had a significant effect on fruit decay during cold storage (p < 0.05). Fruit rotted progressively during storage, except for fruit treated with Ch and Ch+ SA, where no rot was observed up to 45 days (Figure 4). Among all treatments, the control samples had the highest percentage of fruit rot (about 55.6%) after 60 days of storage, while Ch+ SA +Ca, Ch and Ch+ SA had the lowest values (11.6, 25.9-25%) compared to the other treatments (Figure 5). Similarly, SA was reported to be effective in reducing berry drop and rot in grapes at concentrations of 1 and 2 mmol L-1 (Gomes et al., 2021). In addition, SA and Ca alone or in combination increased fruit hardness and delayed softening, resulting in less rot in strawberry fruit (Shafiee et al., 2010).

Equally, elicitors such as SA, Ca and oxalic acid were reported to induce a defence response and reduce decay in other fruits (Tian et al., 2006; Wang et al., 2011; Shen & Shen Yang, 2017). The results of the present study show that the use of Chin in combination with Ca and SA could protect Khesab cv. fruit from decay during cold storage, probably by triggering defence mechanisms in the fruit and maintaining the firmness of the exocarp. Figure 7 illustrates the potentia effect of different preharvest treatments on decay of Khesab cv. fruit during cold storage. Figure 8 shows the possible effect of various preharvest treatments on total contents of Khesab cv. fruit.

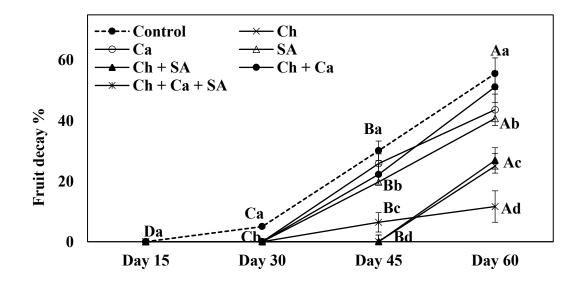
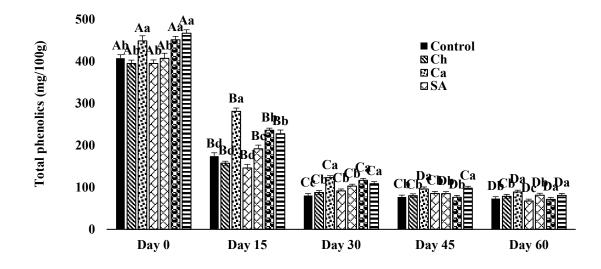
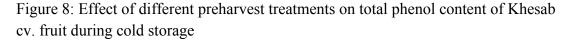


Figure 7: Effect of different preharvest treatments on Khesab cv. fruit decay during cold storage





Note: Values are the mean $(n = 3) \pm SE$. Means with different letters between treatments (small letters) at a different time interval (capital letters) are significantly different at p < 0.05using the LSD test. Ch: chitosan; Ca: calcium chloride; SA: salicylic acid.

4.6 Colour Attributes

Colour is an essential visual attribute of food, especially fresh fruit, which directly affects the attractiveness and acceptance of the product by consumers (FernÁndez-Vázquez et al., 2011). Therefore, the effect of treatments on colour differences was considered when evaluating the quality of fresh fruit of the Khesab variety. Important colour parameters, i.e., ΔE , C*, L*, and h⁰, were determined to follow fruit colour changes during cold storage after elicitor treatments and are presented in Table 4. In general, the colour parameters were significantly (p < 0.05) affected by the Elicitor

application. In addition, colour values gradually decreased with increasing storage time for all fruits. For example, the 1* values of all fruits decreased as storage progressed, with a significant decrease in fruit brightness from 37 to 21 after 60 days. After 30 days, significant differences were found between treatments, as Ch and Ch+ SA showed the least decrease in L*, C*, and h^0 .

After 60 days, the lowest decrease in L* was observed in fruit treated with Ca, followed by SA, Ch and Ch+ SA (Table 4). The same treatments also had lower ΔE values than other treatments, including the control. Similarly, C* was highest in Ch+Ca at the end of the storage period, followed by Ca, Ch, control, Ch+Ca+ SA, Ch+ SA, and SA treated fruits. The L*, C* and h o values were positively correlated with the increase TSS and ripening, (as shown in Table 5), while ΔE was the opposite. The observed delay in colour change of fruits treated with Ca, SA and Ch could be due to the interruption of ethylene action, the unmasking of chlorophyll pigment and the reduction of fruit respiration rate, leading to a slowdown in colour change (Irfan et al., 2013). Similarly, pistachio fruits treated with Ch and SA maintained their colour quality compared to the control (Molamohammadi et al., 2020).

Colour deterioration reactions (browning of date fruits), especially enzymatic reactions, are a naturally occurring phenomenon mainly triggered by the action of peroxidase and polyphenol oxidase during fruit ripening and storage and are associated with a loss of fruit market value (Awad et al., 2011b; Mortazavi et al., 2015). These responses are due to oxidation of phenolic compounds and development of dark brown pigments in date fruits (Awad et al., 2011a). In the present study, the steady decrease in TPC during storage (Figure 7) was positively correlated with the decrease in colour parameters, i.e., h^0 , C*, L* and negatively correlated with ΔE , to varying degrees in

all fruits (Table 4). Thus, based on the present results, it is possible that the application of Ca alone or in combination with Ch and SA delayed the responses to colour change to different degrees and, accordingly, preserved the colour of the fruits.

4.7 Total Phenolic Content of Fruit at Harvest and During Storage

The TPC content of Khesab cv. fruits at harvest and during cold storage is shown in Figure 8. The type of elicitor administered had a significant effect on TPC content at harvest and during cold storage. In general, TPC content decreased with increasing storage time for all fruits. Similarly, phenolic content decreased at different time points from early development stage to maturity and ripening (Awad et al., 2011a). In the present study, Ch+Ca+ SA treated fruits had the highest TPC concentration, which progressively decreased from 486.8mg 100g-1 GAE at day 0 to 66.9mg 100g-1 GAE at day 60, followed by Ca (from 448.5 to 82.8) and Ch (394.6 to 68.5mg 100g-1 GAE) compared to the control and the other treatments.

However, TPC concentration was found to vary (50-400mg 100 g-1) among different date cultivars in several studies (Al-Qurashi & Awad, 2011; Awad et al., 2011b; Mohamed et al., 2014), which could be due to cultivar differences and/or environmental factors. In other fruits, SA has been shown to reduce cold exposure injury in peaches for the duration of cold storage (Wang et al., 2006) due to its ability to induce antioxidant systems and heat shock proteins (Wang et al., 2006). It has been reported that the application of triggers such as chitosan, SA and Ca increases the content of polyphenols in fruits, improving their quality (Ruiz-García & Gómez-Plaza, 2013, Khalil et al., 2022).

In the present study, Ch+ SA +Ca treatment showed low rotting rate during cold storage (Figure 8) and had the lowest rutab content after Ca treatment (Figure 6). In addition, TPC showed a negative correlation with weight loss, rutab content, and decay rate (Table 4), indicating an essential role in date fruit ripening/senescence. Based on these results, preharvest application of Ch, Ch+Ca and Ch+ SA +Ca improved TPC content, which improved fruit quality and shelf life of Khesab cultivar.

				L^*							Hue	Angle		
	Control	Ch	Ca	SA	Ch + SA	Ch + Ca	Ch + Ca + SA	Control	Ch	Ca	SA	Ch + SA	Ch + Ca	Ch + Ca + SA
Day 0	33.1± 0.44b	33.1±0.36b	32.6± 0.58t	o 31.2±0.52b	$\begin{array}{c} 33.44 \pm \\ 0.83b \end{array}$	37.63 ± 0.67a	37.72 ± 0.863	a $27.65 \pm 0.34a$	$\begin{array}{c} 26.03 \pm \\ 0.36b \end{array}$	24.95 ± 0.38c	23.87 ± 0.37c	27.29 ± 0.47a	$24.23 \pm 0.32c$	$27.42 \pm 0.41a$
Day 15	27.4± 0.54b	29.5± 38a	29.8± 0.64a	$27.9 \pm 0.51b$	29.68 ± 0.54a	29.66 ± 0.70a	30.85 ± 0.91a	a $28.88 \pm 0.46b$	30.28 ± 0.45a	26.88 ± 0.31d	$25.39 \pm 0.33d$	27.85 ± 0.39c	$26.12 \pm 0.46d$	$29.94 \pm 0.49a$
Day 30	$25.2\pm0.59b$	26.7± 0.64b	28.4± 0.85a	a 26.0±0.91b	28.50 ± 0.78a	27.08 ± 0.94a	27.43 ± 0.893	a $38.08 \pm 0.51a$	35.93 ± 0.53b	29.99 ± 0.45d	28.80 ± 0.44d	34.17 ± 0.84c	$\begin{array}{c} 34.50 \pm \\ 0.91 \text{c} \end{array}$	$38.74 \pm 0.81a$
Day 45	22.0± 0.32c	23.4± 0.43b	25.2±0.69a	a 22.9± 0.47bc	23.21 ± 0.60b	24.64 ± 0.48a	24.08 ± 0.443	a $38.00 \pm 0.53b$	$\begin{array}{c} 38.30 \pm \\ 0.67 b \end{array}$	35.89 ± 0.74c	34.42 ± 0.68c	37.46 ± 0.73b	$\begin{array}{c} 38.34 \pm \\ 0.94b \end{array}$	$40.61 \pm 0.90a$
Day 60	21.2± 0.52b	21.7± 0.43b	23.7 ± 0.88	a 21.22 ± 0.44b	21.71 ± 0.36b	23.30 ± 0.67a	22.22 ± 0.41b	41.91 ± 0.46a	41.15 ± 0.66a	37.08 ± 0.62c	36.85 ± 0.41c	$39.39 \pm 0.45b$	$\begin{array}{c} 39.39 \pm \\ 0.82b \end{array}$	$42.17 \pm 0.44a$
				Chroma							ΔE			
	Control	Ch	Ca	SA	Ch + SA	Ch + Ca	Ch + Ca + SA	Control	Ch	Ca	SA	Ch + SA	Ch + Ca	Ch + Ca + SA
Day 0	30.67 ± 0.47bc	$\begin{array}{c} 31.77 \pm \\ 0.82b \end{array}$	29.94 ± 0.39c	$28.75 \pm 0.38c$	32.88 ± 0.76b	32.57 ± 0.56b	34.32 ± 0.714a	0e	1.41 ± 0.06	$dd1.67 \pm 0.04d$	13.35 ± 0.13	$b2.25 \pm 0.07c$	5.25 ± 0.10	a 5.87 ± 0.16a
Day 15	$27.36 \pm 0.56c$	$29.77 \pm 0.46b$	29.35 ± 0.44b	28.09 ± 0.41bc	$\begin{array}{c} 32.09 \pm \\ 0.54a \end{array}$	27.55 ± 0.42c	28.55 ± 0.43b	$6.67 \pm 0.14a$	$\begin{array}{c} 3.98 \pm \\ 0.08 \text{bc} \end{array}$	$3.53 \pm 0.08c$	5.85 ± 0.15	$b3.71 \pm 0.09c$:4.71 ± 0.08	b $3.30 \pm 0.05c$
Day 30	$10.41 \pm 0.23d$	12.86 ± 0.21c	15.63 ± 0.24b	13.08 ± 0.25 bc	18.39 ± 0.23a	11.77 ± 0.18c	14.40 ± 0.22b	21.98 ± 0.36a	$19.14 \pm 0.33b$	15.79 ± 0.15c	18.96 ± 0.31b	13.39 ± 0.23d	19.96 ± 0.34b	17.69 ± 0.24b
Day 45	$6.57 \pm 0.12c$	$7.45 \pm 0.11b$	$0.08.70 \pm 0.133$	$a 6.10 \pm 0.11c^{-1}$	$7.32 \pm 0.12b$	5.83 ± 0.14	c $8.91 \pm 0.12a$	$\begin{array}{r} 26.53 \pm \\ 034a \end{array}$	25.33 ± 0.46a	23.46 ± 0.37b	26.65 ± 0.41a	25.48 ± 0.53a	$\begin{array}{c} 26.40 \pm \\ 0.43a \end{array}$	23.89 ± 0.35b
Day 60	$5.15 \pm 0.14a$	$5.70 \pm 0.9a$	5.75 ± 0.11	a 3.69 ± 0.08 b	$4.59 \pm 0.10a$	4.91 ± 0.05	a 4.97 ± 0.09a	$\begin{array}{c} 28.15 \pm \\ 0.57a \end{array}$	$\begin{array}{c} 27.49 \pm \\ 0.72b \end{array}$	26.63 ± 0.51b	29.48 ± 0.64a	$\begin{array}{c} 28.47 \pm \\ 0.69a \end{array}$	28.76 ± 0.47a	27.72 ± 0.42ab

Table 4: Effect of different preharvest treatments on Khesab cv. fruit colour attributes during cold storage

Note: Values are the mean (n = 25) ± SE. Means with different letters in the same row for each colour index are significantly different at p > 0.05 using LSD test. Ch: chitosan; Ca: calcium chloride; SA: salicylic acid.

Traits	TSS	ТРС	Rutab	Decay %	Weight	L*	ΔΕ	Chrom
			%		Loss %			a
ТРС	-0.73							

Rutab	0.74	-0.92						
%	***	***						
Decay	0.61	-0.42	0.56					
%	***	***	***					
Weight	0.52	-0.77	0.65	0.43 ***				
loss %	***	***	***					
L*	-0.67	0.85	-0.91	-0.39	-0.60			
	***	***	***	***	***			
ΔE	0.68	-0.81	0.88	0.47 ***	0.55 ***	-0.96		
	***	***	***			***		
Chroma	-0.63	0.72	-0.85	-0.49	-0.52	0.93	-0.94	
	***	***	***	***	***	***	***	
Hº	-0.67	0.81	-0.89	-0.55	-0.63	0.92	-0.90	0.91
	***	***	***	***	***	***	***	***

Table 5: Pearson's correlation coefficients between some biochemical and physical characteristics of Khesab cv. Fruit

(***) significant at level P = 0.001

4.8 Total Tannin, Flavonoids, and Antioxidants Concentrations at Harvest

The effect of the different elicitors on total tannin (TTC), flavonoids, and antioxidant activity was determined at harvest (Table 6). TTC, TFC, and antioxidant activity affected by the applied elicitors showed significant differences among the different

treatments. Compared with all treatments, fruit treated with Ch+Ca+ SA had the highest TTC and TFC values, 116.5 mg 100 g-1 CE and 91.0 mg 100 g-1 CE, respectively, followed by Ch+Ca (80.0 and 82.7 mg 100 g-1 CE) compared with the control and the other treatments. The lowest levels of these phytochemicals were SA (23.8 and 56.3 mg 100 g-1 CE) and Ch (21 and 73.6 mg 100 g-1 CE). These antioxidant compounds showed higher activity in fruits treated with CH + SA +Ca , as did the radical scavengers ABTS (323.7 mg 100 g-1 TE) and DPPH with the lowest values (IC50 = 1.5 mg mL-1) (Table 6).

The lower IC50 values in the determination of DPPH radical scavenging activities indicate that the extract concentration required to scavenge the DPPH radical decreased by 50%, indicating an improvement in the antioxidant activity of the phenolic compounds found in the fruit. This result is consistent with the low ripening (low rutab content) and decay observed in the same treatment (Figures 6 & 8). In general, the concentration of antioxidants in dates decreases from the early stage (Bisr) to the ripening stage (Awad et al., 2011). These results are in agreement with those found for TPC at harvest (Table 6), highlighting the crucial role of phenolic compounds in the antioxidant capacity of Khesab variety fruits. Moreover, a positive correlation between antioxidant compound content and antioxidant capacity was found in five-date cultivars (Awad et al., 2011a).

In strawberry fruit, preharvest spraying with chitosan increased flavonoids and phenolic compounds up to 2.6-fold compared to the untreated control (Rahman et al., 2018). Application of SA was found to increase antioxidant compounds in table grapes (Gomes et al., 2021). Comparable results were also reported by Mohamed et al. (2014). They found that preharvest spraying with SA significantly increased DPPH

scavenging activity at the bisr stage of date fruit compared to control fruit. In date, as a climacteric fruit, oxidative stress during ripening is thought to be responsible for the reduction of antioxidant compounds (Figure 7) at the bisr and/or rutab stages as a result of the decrease in free radical scavenging ability (Mohamed et al., 2014). Therefore, more reactive oxygen species (ROS), including H2O2 and superoxide, may develop and be involved in the ripening/senescence of date fruit and other fruits due to the development of free radicals (Ferrer et al., 2005, Khalil et al., 2022). According to the above results, preharvest application of Ch+ SA +Ca increased the antioxidant compound content and antioxidant activity, which improved the quality and shelf life of Khesab cv. fruits possibly by reducing harmful radicals.

	TPC	TTC	TFC	(DPPH) IC50	ABTS
	(mg/100 gm)	(mg/100 gm)	(mg/100 g)	(mg/mL)	(mg/g)
Control	$407.4 \pm 12.2c$	73.9±0.75c	71.5 ± 0.34 bc	$1.7\pm0.03b$	$1.9\pm0.07b$
Ch	$394.6 \pm 8.89c$	$42.0 \pm 0.43e$	87.8± 0.42b	3.3 ± 0.10a	$1.6 \pm 0.11c$
Ca	$448.5 \pm 11.3b$	61.9 ± 0.68 d	$67.9 \pm 0.22c$	$1.7 \pm 0.06b$	$1.5 \pm 0.09c$
SA	$394.9 \pm 10.7c$	$47.8 \pm 0.26e$	$66.4 \pm 0.31c$	$2.1\pm0.07b$	1.3 ± 0.17 d
Ch + SA	$407.2 \pm 13.2c$	$59.5 \pm 0.45 d$	$86.6\pm0.49b$	1.8 ± 0.05 ab	$1.9 \pm 0.18b$
Ch + Ca	$451.1 \pm 13.4b$	$80.7\pm0.64b$	$82.9\pm0.54b$	1.8 ± 0.04 ab	$1.2 \pm 0.12d$
Ch + Ca + SA	486.8 ± 12.7a	116.2 ± 0.73 a	91.4 ± 0.41a	$1.5 \pm 0.04b$	3.2 ± 0.13a

Table 6: Effect of different preharvest treatments on Khesab cv. fruit phytochemical analysis at harvest

Note: Values are the mean $(n = 3) \pm SE$ for the phytochemical analysis. Means with different letters in the same column are significantly different at p > 0.05 using LSD test. TPC = Total Phenolic Content; TTC = Total Tannin Content; TFC = Total flavonoid Content. Ch: chitosan; Ca: calcium chloride; SA: salicylic acid.

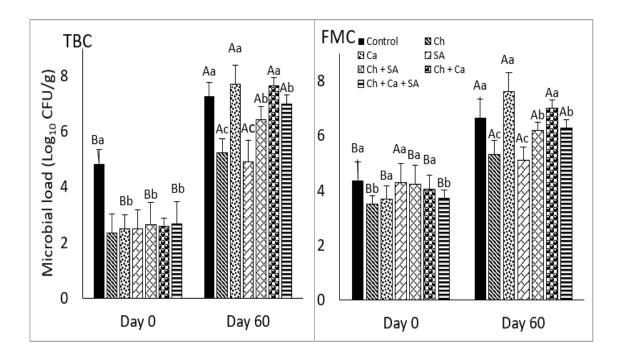
4.9 Microbiological Quality of Date Fruit

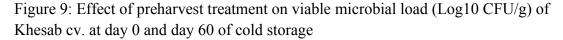
The microbial load (Log10 CFU g-1) of Khesab cv. samples at harvest and at the end of cold storage is shown in Figure 9. The analysis was performed for only two days, at day 0 and day 60. In general, total bacterial count (TBC) and fungal/mould count

(FMC) at harvest and at the end of storage showed significant variations influenced by the application of triggers. For example, date fruit treated with Ch, SA, Ca, Ch+Ca, Ch+ SA and Ch+Ca+ SA had lower TB counts than the control group at day 0, while FMC counts were significantly lower in fruit treated with Ch, Ca and Ch+Ca+ SA compared to the other treatments and the control group.

After 60 days, SA and Ch-treated fruits showed lower FMC counts, followed by Ch+ SA and Ch+Ca+ SA compared to the control group and the other treatments, while the same treatments showed significantly lower FMC counts than the control group and the other treatments (Figure 9). Consistently, different elicitors such as SA showed the best results in eliciting defence responses and minimising microbial infestation and rot in pear fruit (Tian et al., 2006). In addition, chitosan has been shown to stimulate plant defences against various pathogens, including fungi and bacteria (Romanazzi et al., 2018, Ahmed et al., 2021a). Moreover, treatment with Ch and SA either separately or in combination drastically reduced bacterial and fungal growth in pistachio fruits (Molamohammadi et al., 2020).

Based on the present results, the combination treatment with different elicitors showed better antimicrobial activity than other treatments. Moreover, the same treatment showed the highest antioxidant activity and the lowest percentage of decay, which explains the results of microbial analysis (Irfan et al., 2013). Thus, the increase in antioxidant activity and phenolics by treatment with different elicitors alone or in combination may induce defence mechanisms that lead to resistance to pathogens and prolong fruit shelf life, as has been reported in other fruits (Wang et al., 2011; Sathiyabama et al., 2014; Romanazzi et al., 2018).





Note: Values are the mean $(n = 3) \pm SE$. Means with different letters between treatments (small letters) at a different time interval (capital letters) are significantly different at p < 0.05 using the LSD test. TBC = Total Bacterial Count, FMC = Yeast and Mold Count. Ch: chitosan;

Ca: calcium chloride; SA: salicylic acid.

Chapter 5: Summary and Conclusions

This study showed that the application of different combinations of elicitors improved the biochemical properties of fresh fruits of Khesab cv. throughout the ripening and storage period. The synergistic effect of elicitors was found when different treatments were combined; Ch+Ca and Ch+Ca+ SA and Ch+ SA treated fruits in harvested Bisr date fruits showed reduced rot and microbial load. At the end of cold storage, Ca followed by Ch+Ca+ SA had the highest TPC concentration as they delayed natural ripening. Moreover, antioxidant activity was highest in Ch+ SA followed by Ch+ SA +Ca and Ch compared to the other treatments.

These results suggest that the delay in ripening and reduction in fruit decay may be due to the regulating effect of the sprayed elicitors on the ripening process and thus delaying senescence of fresh fruit of Khesab cultivar. Based on the results of this experiment, (Ch+ SA and Ch+ SA +Ca) may be considered suitable for large-scale application and should be tested on different dates and other fruits. The use of natural compounds can be an efficient, cost-effective and natural way to improve the quality and marketability of date fruits and protect them from a variety of decaying microbes.

References

- Abd Elwahab, S. M., Abd Allatif, A. M., Farid, M. A., & Soliman, S. M. (2019). Effect of safe post-harvest alternatives on quality and storage life of "barhi" date palm. Plant Archives, 19(2), 3937-3945.
- Abdel-Hamid, A. N. (2020). Changes in Post-harvest Quality of Menthe and Sage fresh Cut Herbs Treated with Citric acid, Salicylic Acid and Chitosan under Cold storage. *Middle East Journal of Agriculture Research*, 9(1), 134-148.
- Abu-Shama, H. S., Abou-Zaid, F. O. F., & El-Sayed, E. Z. (2020). Effect of using edible coatings on fruit quality of Barhi date cultivar. *Scientia Horticulturae*, 265, 109262.
- Ahmed, I. A., Ahmed, A. W. K., & Robinson, R. K. (1995). Chemical composition of date varieties as influenced by the stage of ripening. *Food chemistry*, 54(3), 305-309.
- Ahmed, Z. F., Al Shaibani, F. Y., Kaur, N., Maqsood, S., & Schmeda-Hirschmann, G. (2021b). Improving fruit quality, bioactive compounds, and storage life of date palm (*Phoenix dactylifera* L., cv. Barhi) using natural elicitors. *Horticulturae*, 7, 293.
- Ahmed, Z. F., Alblooshi, S. S., Kaur, N., Maqsood, S., & Schmeda-Hirschmann, G. (2021a). Synergistic effect of preharvest spray application of natural elicitors on storage life and bioactive compounds of date palm (*Phoenix dactylifera* L., cv. Khesab). *Horticulturae*, 7,145.
- Ahmed, Z. F., Kaur, N., Maqsood, S., & Schmeda-Hirschmann, G. (2022b). Preharvest applications of chitosan, salicylic acid, and calcium chloride have a synergistic effect on quality and storability of date palm fruit (*Phoenix dactylifera* L.). *HortScience*, 57 (3), 422-430.
- Ahmed, Z. F. R & J. P., Palta. (2010) Lysophosphatidylethanolamine, a natural phospholipid, may retard senescence and improve the shelf life of banana fruit. *HortScience*. *45*, S66-S66.
- Ahmed, Z. F. R & J.P., Palta. (2011) Hormone-like effect of a natural lipid, lysophosphatidylethanolamine, can mitigate calcium deficiency injury in potato shoot cultures. HortScience. 46, S196-S197.
- Ahmed, Z.F., Kaur, N. and Hassan, F. E. (2022a). Ornamental date palm and sidr trees: fruit elements composition and concerns regarding consumption. *International. Journal of Fruit Science*, 22, 17-34.

- Ahmed, Z.F.R., Alnuaimi, A.K.H., Askri, A., & Tzortzakis, N. (2021c). Evaluation of lettuce (*Lactuca sativa* L.) production under hydroponic system: nutrient solution derived from fish waste vs. inorganic nutrient solution. *Horticulturae*, 7, 292.
- Al-Alawi, R. A., Al-Mashiqri, J. H., Al-Nadabi, J. S., Al-Shihi, B. I., & Baqi, Y.
 (2017). Date palm tree (Phoenix dactylifera L.): natural products and therapeutic options. *Frontiers in plant science*, *8*, 845.
- Aleid, S. M., & Saikhan, M. S. A. L. (2017). Effect of permeable modified atmosphere packaging on quality and shelf life of fresh Khenaizy dates stored at low temperature. *Journal of Food and Nutrition Research*, 5, 503-509.
- Al-Eryani-Raqeeb, A., Mahmud, T. M. M., Syed Omar, S. R., Mohamed Zaki, A. R., & Al-Eryani, A. R. (2009). Effects of calcium and chitosan treatments on controlling anthracnose and postharvest quality of papaya (Carica papaya L.). *International Journal of Agricultural Research*, 4(2), 53-68.
- Al-Farsi, M. A., & Lee, C. Y. (2008). Optimization of phenolics and dietary fiber extraction from date seeds. *Food Chemistry*, 108(3), 977-985.
- Al-Harrasi, A., Rehman, N. U., Hussain, J., Khan, A. L., Al-Rawahi, A., Gilani, S. A., Al-Broumi, M. & Ali, L. (2014). Nutritional assessment and antioxidant analysis of 22 date palm (Phoenix dactylifera) varieties growing in Sultanate of Oman. *Asian Pacific Journal of Tropical Medicine*, 7, S591-S598.
- Al-Mssallem, I. S., Hu, S., Zhang, X., Lin, Q., Liu, W., Tan, J., ..., & Yu, J. (2013). Genome sequence of the date palm *Phoenix dactylifera* L. *Nature Communications*, 4(1), 1-9.
- Al-Mssallem, M. Q., Alqurashi, R. M., & Al-Khayri, J. M. (2020). Bioactive compounds of date palm (Phoenix dactylifera L.). *Bioactive Compounds in Underutilized Fruits and Nuts*, 91-105.
- Al-Qarawi, A. A., Ali, B. H., Al-Mougy, S. A., & Mousa, H. M. (2003). Gastrointestinal transit in mice treated with various extracts of date (*Phoenix dactylifera* L.). *Food and Chemical Toxicology*, 41(1), 37-39.
- Alqarni, S., & Bazzi, M. D. (2019). Evaluating the *Phoenix dactylifera* L. biochemical changes associated with its maturation. *Journal of Agricultural Science*, 11(3), 614-624.
- Al-Qurashi, A. D., & Awad, M. A. (2011). Quality characteristics of bisir
 'Barhee' dates during cold storage as affected by postharvest dipping in gibberellic acid, naphthaleneacetic acid and benzyladenine. *Fruits*, 66(5), 343-352.

- Al-Qurashi, A. D., & Awad, M. A. (2015). Postharvest chitosan treatment affects quality, antioxidant capacity, antioxidant compounds and enzymes activities of 'El-Bayadi'table grapes after storage. *Scientia Horticulturae*, 197, 392-398.
- Alsawmahi, O. N., Al-Juhaimi, F., Alhamdan, A. M., Ghafoor, K., Adiamo, O. Q., ..., & Alashmawe, N. (2018). Phenolic, tannin, antioxidant, color, and sensory attributes of Barhi date (Phoenix dactylifera) fruit stored in modified atmosphere packages. *Journal of Food Biochemistry*, 42(5), e12576.
- Al-Yahyai, R., & Al-Kharusi, L. (2012). Physical and chemical quality attributes of freeze-stored dates. *International Journal of Agriculture and Biology*, 14(1).
- Amira, E. A., Behija, S. E., Beligh, M., Lamia, L., Manel, I., Mohamed, H., & Lotfi, A. (2012). Effects of the ripening stage on phenolic profile, phytochemical composition and antioxidant activity of date palm fruit. *Journal of agricultural and food chemistry*, 60(44), 10896-10902.
- Arnon, H., Zaitsev, Y., Porat, R., & Poverenov, E. (2014). Effects of carboxymethyl cellulose and chitosan bilayer edible coating on postharvest quality of citrus fruit. *Postharvest Biology and Technology*, 87, 21-26.
- Atia, A., Abdelkarim, D., Younis, M., & Alhamdan, A. (2018). Effects of calcium chloride and salicylic acid postharvest treatments on the quality of Khalal Barhi dates at different ripening levels during cold storage. *Journal of Food Measurement and Characterization*, 12(2), 1156-1166.
- Awad, M. A., & Al-Qurashi, A. D. (2012). Gibberellic acid spray and bunch bagging increase bunch weight and improve fruit quality of 'Barhee'date palm cultivar under hot arid conditions. *Scientia Horticulturae*, 138, 96-100.
- Awad, M. A., Al-Qurashi, A. D., & Mohamed, S. A. (2011a). Antioxidant capacity, antioxidant compounds and antioxidant enzyme activities in five date cultivars during development and ripening. *Scientia Horticulturae*, 129(4), 688-693.
- Awad, M. A., Al-Qurashi, A. D., & Mohamed, S. A. (2011b). Biochemical changes in fruit of an early and a late date palm cultivar during development and ripening. *International Journal of Fruit Science*, 11(2), 167-183.
- Badawy, I. F., Abou-Zaid, E. A., & Hussein, E. M. (2019). Cracking and fruit quality of "Manfalouty" pomegranate as affected by pre-harvest of chitosan, calcium chloride and gibbrellic acid spraying. *Middle East Journal of Agriculture*, 8(3), 873-882.
- Baenas, N., García-Viguera, C., & Moreno, D. A. (2014). Elicitation: a tool for enriching the bioactive composition of foods. *Molecules*, 19(9), 13541-13563.
- Baldwin, E. A., Hagenmaier, R., & Bai, J. (Eds.). (2011). *Edible Coatings and Films to Improve Food Quality* (variuos pages). London: CRC Press.

- Baloch, M. K., Saleem, S. A., Baloch, A. K., & Baloch, W. A. (2006). Impact of controlled atmosphere on the stability of Dhakki dates. *LWT-food Science & Technology*, 39(6), 671-676.
- Barreto, T. A., Andrade, S. C. A., Maciel, J. F., Arcanjo, N. M. O., Madruga, M. S., Meireles, B., Cordeiro, Â. M. T., Souza, E. L. & Magnani, M. (2016). A chitosan coating containing essential oil from Origanum vulgare L. to control postharvest mold infections and keep the quality of cherry tomato fruit. *Frontiers in Microbiology*, 7, 1724.
- Bautista-Baños, S., Hernandez-Lauzardo, A. N., Velazquez-Del Valle, M. G., Hernández-López, M., Barka, E. A., Bosquez-Molina, E., & Wilson, C. L. (2006). Chitosan as a potential natural compound to control pre and postharvest diseases of horticultural commodities. *Crop Protection*, 25(2), 108-118.
- Bautista-Baños, S., Ventura-Aguilar, R. I., Correa-Pacheco, Z., & Corona-Rangel, M. L. (2017). Chitosan: a versatile antimicrobial polysaccharide for fruit and vegetables in postharvest-a review. *Revista Chapingo. Serie horticultura*, 23(2), 103-122.
- Ben-Shalom, N., Ardi, R., Pinto, R., Aki, C., & Fallik, E. (2003). Controlling gray mould caused by Botrytis cinerea in cucumber plants by means of chitosan. *Crop Protection*, 22(2), 285-290.
- Bentebba, F. Z., Zineb, G., Saidi, M., & Bensaci, C. (2020). Effects of development and ripening stage on phytochemical compositions, antioxidant and antibacterial activities of date palm fruits. *Asian Journal of Chemistry*, *32*(2), 415-419.
- Bentrad, N., & Hamida-Ferhat, A. (2020). Date palm fruit (*Phoenix dactylifera*): Nutritional values and potential benefits on health. In *The Mediterranean Diet* (pp. 239-255). Academic Press.
- Biglari, F., AlKarkhi, A. F., & Easa, A. M. (2009). Cluster analysis of antioxidant compounds in dates (Phoenix dactylifera): Effect of long-term cold storage. *Food Chemistry*, 112(4), 998-1001.
- Bill, M., Sivakumar, D., Korsten, L., & Thompson, A. K. (2014). The efficacy of combined application of edible coatings and thyme oil in inducing resistance components in avocado (Persea americana Mill.) against anthracnose during post-harvest storage. *Crop Protection*, 64, 159-167.
- Chao, C. T., & Krueger, R. R. (2007). The date palm (Phoenix dactylifera L.): overview of biology, uses, and cultivation. *HortScience*, *42*(5), 1077-1082.
- Chawla, S., Devi, R., & Jain, V. (2018). Changes in physicochemical characteristics of guava fruits due to chitosan and calcium chloride treatments during storage. *Journal of Pharmacognosy and Phytochemistry*, 7(3), 1035-1044.

- Chen, J., Zou, X., Liu, Q., Wang, F., Feng, W., & Wan, N. (2014). Combination effect of chitosan and methyl jasmonate on controlling Alternaria alternata and enhancing activity of cherry tomato fruit defense mechanisms. *Crop Protection*, 56, 31-36.
- Chen, Y., Sun, J., Lin, H., Lin, M., Lin, Y., Wang, H., & Hung, Y. C. (2020). Salicylic acid treatment suppresses Phomopsis longanae Chi-induced disease development of postharvest longan fruit by modulating membrane lipid metabolism. *Postharvest Biology and Technology*, 164, 111168.
- Chong, J. X., Lai, S., & Yang, H. (2015). Chitosan combined with calcium chloride impacts fresh-cut honeydew melon by stabilising nanostructures of sodiumcarbonate-soluble pectin. *Food Control*, 53, 195-205.
- Cindi, M. D., Shittu, T., Sivakumar, D., & Bautista-Baños, S. (2015). Chitosan boehmite-alumina nanocomposite films and thyme oil vapour control brown rot in peaches (Prunus persica L.) during postharvest storage. *Crop Protection*, 72, 127-131.
- Cissé, M., Polidori, J., Montet, D., Loiseau, G., & Ducamp-Collin, M. N. (2015). Preservation of mango quality by using functional chitosan-lactoperoxidase systems coatings. *Postharvest Biology and Technology*, 101, 10-14.
- Dransfield, J., Uhl, N., Asmussen, C. B., Baker, W. J., Harley, M. M., & Lewis, C. E. (2005). A new phylogenetic classification of the palm family *Arecaceae*. *Kew Bulletin*, 559-569.
- Duran, M., Aday, M. S., Zorba, N. N. D., Temizkan, R., Büyükcan, M. B., & Caner, C. (2016). Potential of antimicrobial active packaging 'containing natamycin, nisin, pomegranate and grape seed extract in chitosan coating'to extend shelf life of fresh strawberry. *Food and Bioproducts Processing*, *98*, 354-363.
- Duthie, G. G., Gardner, P. T., & Kyle, J. A. (2003). Plant polyphenols: are they the new magic bullet?. *Proceedings of the Nutrition Society*, *62*(3), 599-603.
- El Ghaouth, A., Ponnampalam, R., Castaigne, F., & Arul, J. (1992). Chitosan coating to extend the storage life of tomatoes. *HortScience*, *27*(9), 1016-1018.
- El-Badawy, H. E. M. (2012). Effect of chitosan and calcium chloride spraying on fruits quality of Florida Prince peach under cold storage. *Research Journal of Agriculture and Biological Sciences*, 8(2), 272-281.
- Elbarbary, A. M., & Mostafa, T. B. (2014). Effect of γ-rays on carboxymethyl chitosan for use as antioxidant and preservative coating for peach fruit. *Carbohydrate Polymers*, *104*, 109-117.

- Ennab, H. A., El-Shemy, M. A., & Alam-Eldein, S. M. (2020). Salicylic acid and putrescine to reduce post-harvest storage problems and maintain quality of Murcott Mandarin fruit. *Agronomy*, 10(1), 115.
- Feliziani, E., Landi, L., & Romanazzi, G. (2015). Preharvest treatments with chitosan and other alternatives to conventional fungicides to control postharvest decay of strawberry. *Carbohydrate Polymers*, 132, 111-117.
- FernÁndez-VÁzquez, R., Stinco, C.M., MelÉndez-MartÍnez, A.J, Heredia, F.J., Vicario, I.M. (2011). Visual and instrumental evaluation of orange juice color: A consumers'preference study. *Journal of Sensory Studies*, 26(6), 436-444.
- Ferrer, A., Remón, S., Negueruela, A. I., & Oria, R. (2005). Changes during the ripening of the very late season Spanish peach cultivar Calanda: feasibility of using CIELAB coordinates as maturity indices. *Scientia Horticulturae*, 105(4), 435-446.
- Frankel, E. N., German, J. B., Kinsella, J. E., Parks, E., & Kanner, J. (1993). Inhibition of oxidation of human low-density lipoprotein by phenolic substances in red wine. *The Lancet*, 341(8843), 454-457.
- Gantait, S., El-Dawayati, M. M., Panigrahi, J., Labrooy, C., & Verma, S. K. (2018). The retrospect and prospect of the applications of biotechnology in Phoenix dactylifera L. *Applied microbiology and biotechnology*, *102*(19), 8229-8259.
- Gayed, A. A. N. A., Shaarawi, S. A. M. A., Elkhishen, M. A., & Elsherbini, N. R. M. (2017). Pre-harvest application of calcium chloride and chitosan on fruit quality and storability of 'Early Swelling'peach during cold storage. *Ciência e Agrotecnologia*, 41, 220-231.
- Gol, N. B., Patel, P. R., & Rao, T. R. (2013). Improvement of quality and shelf-life of strawberries with edible coatings enriched with chitosan. *Postharvest Biology* and Technology, 85, 185-195.
- Gomes, E. P., Borges, C. V., Monteiro, G. C., Belin, M. A. F., Minatel, I. O., ..., & Lima, G. P. P. (2021). Preharvest salicylic acid treatments improve phenolic compounds and biogenic amines in 'Niagara Rosada'table grape. *Postharvest Biology & Technology*, 176, 111505.
- Guerra, I. C. D., de Oliveira, P. D. L., Santos, M. M. F., Lúcio, A. S. S. C., Tavares, J. F., ..., & de Souza, E. L. (2016). The effects of composite coatings containing chitosan and Mentha (piperita L. or x villosa Huds) essential oil on postharvest mold occurrence and quality of table grape cv. Isabella. *Innovative Food Science & Emerging Technologies*, 34, 112-121.

- Guo, C. E., He, Y., Cui, Q., & Li, W. (2020). Influence of preharvest calcium spray and postharvest chitosan coating methods on quality of Chinese dwarf cherry (Cerasus humilis (Bge.) Sok) fruits during cold storage. *The Journal of Horticultural Science and Biotechnology*, 95(6), 773-781.
- Habib, H. M., & Ibrahim, W. H. (2011). Nutritional quality of 18 date fruit varieties. International Journal of Food Sciences and Nutrition, 62(5), 544-551.
- Hadrami, I. E., & Hadrami, A. E. (2009). Breeding date palm. In *Breeding Plantation Tree Crops: Tropical Species* (pp. 191-216). New York: Springer.
- Haider, M. Khan, I. Naqvi, S. Jaskani, M. Khan, R. Nafees, M., & Pasha, I. (2013). Fruit developmental stages effects on biochemical attributes in date palm. *Pakistan Journal of Agricultural Sciences*, 50(4), 577-583.
- Hassan, O., & Chang, T. (2017). Chitosan for eco-friendly control of plant disease. *Asian Journal of Plant Pathology*, *11*, 53-70.
- Hassni, M., El Hadrami, A., Daayf, F., Barka, E., & El Hadrami, I. (2004). Chitosan, Antifungal Product against" Fusarium oxysporum" f. sp." albedinis" and Elicitor of Defence Reactions in Date Palm Roots. *Phytopathologia Mediterranea*, 43(2), 195-204.
- Hazbavi, I., Khoshtaghaza, M. H., Mostaan, A., & Banakar, A. (2015). Effect of postharvest hot-water and heat treatment on quality of date palm (cv. Stamaran). *Journal of the Saudi Society of Agricultural Sciences*, *14*(2), 153-159.
- He, G., Chen, X., Yin, Y., Zheng, H., Xiong, X., & Du, Y. (2011). Synthesis, characterization and antibacterial activity of salicyloyl chitosan. *Carbohydrate Polymers*, 83(3), 1274-1278.
- He, Y., Bose, S. K., Wang, W., Jia, X., Lu, H., & Yin, H. (2018). Pre-harvest treatment of chitosan oligosaccharides improved strawberry fruit quality. *International journal of molecular sciences*, 19(8), 2194.
- Hernández-Muñoz, P., Almenar, E., Del Valle, V., Velez, D., & Gavara, R. (2008). Effect of chitosan coating combined with postharvest calcium treatment on strawberry (Fragaria× ananassa) quality during refrigerated storage. *Food Chemistry*, 110(2), 428-435.
- Hinkaew, J., Aursalung, A., Sahasakul, Y., Tangsuphoom, N., & Suttisansanee, U. (2021). A comparison of the nutritional and biochemical quality of date palm fruits obtained using different planting techniques. *Molecules*, 26(8), 2245.
- Huang, Z., Li, J., Zhang, J., Gao, Y., & Hui, G. (2017). Physicochemical properties enhancement of Chinese kiwi fruit (Actinidia chinensis Planch) via chitosan coating enriched with salicylic acid treatment. *Journal of Food Measurement and Characterization*, 11(1), 184-191.

- Irfan, P. K., Vanjakshi, V., Prakash, M. K., Ravi, R., & Kudachikar, V. B. (2013). Calcium chloride extends the keeping quality of fig fruit (Ficus carica L.) during storage and shelf-life. *Postharvest biology and technology*, 82, 70-75.
- Ismail, B., Haffar, I., Baalbaki, R., & Henry, J. (2008). Physico-chemical characteristics and sensory quality of two date varieties under commercial and industrial storage conditions. *LWT-Food Science and Technology*, 41(5), 896-904.
- Johnson, D. V. (2012). Enhancement of date palm as a source of multiple products: Examples from other industrialized palms. *Emirates Journal of Food and Agriculture*, 24 (5), 408-414.
- Kader, A. A., & Hussein, A. M. (2009). Harvesting and postharvest handling of dates. *ICARDA, Aleppo, Syria*, 4, 15.
- Kassem, H. A., Al-Obeed, R. S., & Ahmed, M. A. (2010, March). Extending harvest season, improving fruit quality and shelf life of Barhee'date palm by preharvest sprays. In *IV International Date Palm Conference 882* (pp. 147-154).
- Katiyar, D., Hemantaranjan, A., Singh, B., & Bhanu, A. N. (2014). A future perspective in crop protection: Chitosan and its oligosaccharides. *Advances in Plants & Agriculture Research*, 1(1), 1-8.
- Kaur, M., Kalia, A., & Thakur, A. (2017). Effect of biodegradable chitosan–rice-starch nanocomposite films on post-harvest quality of stored peach fruit. *Starch-Stärke*, 69(1-2), 1600208.
- Khadiga, A. B. (1993). Effect of three growth retardants on onion (Allium cepa L.). III. Effect on cytological behaviors' come vegetative characters and their residual effect on C. *Egyptian Journal of Applied Science*, 8(3), 259.
- Khalifa, I., Barakat, H., El-Mansy, H. A., & Soliman, S. A. (2016). Improving the shelf-life stability of apple and strawberry fruits applying chitosan-incorporated olive oil processing residues coating. *Food Packaging and Shelf Life*, *9*, 10-19.
- Khalifa, I., Barakat, H., El-Mansy, H. A., & Soliman, S. A. (2017). Preserving apple (Malus domestica var. Anna) fruit bioactive substances using olive wastes extract-chitosan film coating. *Information Processing in Agriculture*, 4(1), 90-99.
- Khalil, H. A., El-Ansary, D. O., & Ahmed, Z. F. R. (2022). Mitigation of Salinity Stress on Pomegranate (Punica granatum L. cv. Wonderful) Plant Using Salicylic Acid Foliar Spray. *Horticulturae*, 8 (5), 375
- Khaliq, G., Mohamed, M. T. M., Ding, P., Ghazali, H. M., & Ali, A. (2016). Storage behaviour and quality responses of mango (Mangifera indica L.) fruit treated with chitosan and gum arabic coatings during cold storage conditions. *International Food Research Journal*, 23.

- Khatri, D., Panigrahi, J., Prajapati, A., & Bariya, H. (2020). Attributes of Aloe vera gel and chitosan treatments on the quality and biochemical traits of post-harvest tomatoes. *Scientia Horticulturae*, *259*, 108837.
- Kim, D. O., Chun, O. K., Kim, Y. J., Moon, H. Y., & Lee, C. Y. (2003). Quantification of polyphenolics and their antioxidant capacity in fresh plums. *Journal of Agricultural & Food Chemistry*, 51(22), 6509-6515.
- Kim, S. (2018). Competitive biological activities of chitosan and its derivatives: antimicrobial, antioxidant, anticancer, and anti-inflammatory activities. *International Journal of Polymer Science*, 2018.
- Kumar, D., Mishra, D. S., Chakraborty, B., & Kumar, P. (2013). Pericarp browning and quality management of litchi fruit by antioxidants and salicylic acid during ambient storage. *Journal of food science and technology*, 50(4), 797-802.
- Kumar, P., Sethi, S., Sharma, R. R., Srivastav, M., & Varghese, E. (2017). Effect of chitosan coating on postharvest life and quality of plum during storage at low temperature. *Scientia Horticulturae*, 226, 104-109.
- Kumarihami, H. M. P. C., Cha, G. H., Kim, J. G., Kim, H. L., Lee, M., Kwack, Y. B., Cho, J. G & Kim, J. (2020). Effect of preharvest Ca-Chitosan application on postharvest quality of 'Garmrok'kiwifruit during cold storage. *Horticultural Science and Technology*, 38(2), 239-248.
- Kumarihami, H. M., Kim, J. G., Kim, Y. H., Lee, M., Lee, Y. S., Kwack, Y. B., & Kim, J. (2021). Preharvest Application of Chitosan Improves the Postharvest Life of 'Garmrok'Kiwifruit through the Modulation of Genes Related to Ethylene Biosynthesis, Cell Wall Modification, and Lignin Metabolism. *Foods*, 10(2), 373.
- Li, Y., Rokayya, S., Jia, F., Nie, X., Xu, J., Elhakem, A., Almatrafi, M., Benajiba, N. & Helal, M. (2021). Shelf-life, quality, safety evaluations of blueberry fruits coated with chitosan nano-material films. *Scientific Reports*, 11(1), 1-10.
- Lin, K. H., Lin, T. Y., Wu, C. W., & Chang, Y. S. (2021). Protective Effects of Salicylic Acid and Calcium Chloride on Sage Plants (Salvia officinalis L. and Salvia elegans Vahl) under High-Temperature Stress. *Plants*, 10(10), 2110.
- Liu, K., Yuan, C., Chen, Y., Li, H., & Liu, J. (2014). Combined effects of ascorbic acid and chitosan on the quality maintenance and shelf life of plums. *Scientia Horticulturae*, 176, 45-53.
- Liu, X., Ren, J., Zhu, Y., Han, W., Xuan, H., & Ge, L. (2016). The preservation effect of ascorbic acid and calcium chloride modified chitosan coating on fresh-cut apples at room temperature. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 502, 102-106.

- Luo, Z., Chen, C., & Xie, J. (2011). Effect of salicylic acid treatment on alleviating postharvest chilling injury of 'Qingnai'plum fruit. *Postharvest biology and technology*, 62(2), 115-120.
- Maan, A.A., Ahmed, Z.F.R., Khan, M.K.I., Riaz, A. & Nazir, A., (2021). Aloe vera gel, an excellent base material for edible films and coatings. *Trends in Food Science and Technology*, 116, 329-341.
- Mattar, M. A., Soliman, S. S., & Al-Obeed, R. S. (2021). Effects of various quantities of three irrigation water types on yield and fruit quality of 'succary'date palm. *Agronomy*, 11(4), 796.
- Meng, X., & Tian, S. (2009). Effects of preharvest application of antagonistic yeast combined with chitosan on decay and quality of harvested table grape fruit. *Journal of the Science of Food and Agriculture*, 89(11), 1838-1842.
- Mohamed, M. A. A., El-Khalek, A. F. A., & Elmehrat, H. G. (2017). Preharvest applications of potassium silicate, chitosan and calcium chloride to improve mango cv. 'Zibda' fruit quality and storability. *Egyptian Journal of Horticulture*, *44*, 17-32.
- Mohamed, S. A., Awad, M. A., & Al-Qurashi, A. D. (2014). Antioxidant activity, antioxidant compounds, antioxidant and hydrolytic enzymes activities of 'Barhee'dates at harvest and during storage as affected by pre-harvest spray of some growth regulators. *Scientia Horticulturae*, 167, 91-99.
- Mohammadi, A., Hashemi, M., & Hosseini, S. M. (2015). Nanoencapsulation of Zataria multiflora essential oil preparation and characterization with enhanced antifungal activity for controlling Botrytis cinerea, the causal agent of gray mould disease. *Innovative food science & emerging technologies*, *28*, 73-80.
- Molamohammadi, H., Pakkish, Z., Akhavan, H. R., & Saffari, V. R. (2020). Effect of salicylic acid incorporated chitosan coating on shelf life extension of fresh in-hull pistachio fruit. *Food and Bioprocess Technology*, 13(1), 121-131.
- Moreno-Escamilla, J. O., Alvarez-Parrilla, E., Laura, A., Núñez-Gastélum, J. A., González-Aguilar, G. A., & Rodrigo-García, J. (2018). Effect of elicitors in the nutritional and sensorial quality of fruits and vegetables. In *Preharvest Modulation of Postharvest Fruit and Vegetable Quality* (pp. 71-91). London: Academic Press.
- Mortazavi, S. M. H., Azizollahi, F., & Moalemi, N. (2015). Some quality attributes and biochemical properties of nine Iranian date (Phoenix dactylifera L.) cultivars at different stages of fruit development. *International Journal of Horticultural Science and Technology*, 2(2), 161-171.

- Munhuweyi, K., Caleb, O. J., Lennox, C. L., van Reenen, A. J., & Opara, U. L. (2017). In vitro and in vivo antifungal activity of chitosan-essential oils against pomegranate fruit pathogens. *Postharvest Biology and Technology*, 129, 9-22.
- Muñoz, Z., Moret, A., & Garcés, S. (2009). Assessment of chitosan for inhibition of Colletotrichum sp. on tomatoes and grapes. *Crop protection*, *28*(1), 36-40.
- Nguyen, D. H. H., & Nguyen, H. V. H. (2021). Effects of storage temperature on postharvest physico-chemical attributes of nano-chitosan coated strawberry (Fragaria× ananassa Duch.). *Journal of Horticulture and Postharvest Research*, 4, 101-114.
- Nia, A. E., Taghipour, S., & Siahmansour, S. (2021). Pre-harvest application of chitosan and postharvest Aloe vera gel coating enhances quality of table grape (Vitis vinifera L. cv. 'Yaghouti') during postharvest period. *Food Chemistry*, 347, 129012.
- Nirupama, P., Gol, N. B., & Rao, T. R. (2010). Effect of post-harvest treatments on physicochemical characteristics and shelf life of tomato (*Lycopersicon esculentum* Mill.) fruits during storage. *American-Eurasian Journal of Agricultural and Environmental Sciences*, 9(5), 470-479.
- Ochoa-Velasco, C. E., & Guerrero-Beltrán, J. Á. (2014). Postharvest quality of peeled prickly pear fruit treated with acetic acid and chitosan. *Postharvest Biology and Technology*, 92, 139-145.
- Pasquariello, M. S., Di Patre, D., Mastrobuoni, F., Zampella, L., Scortichini, M., & Petriccione, M. (2015). Influence of postharvest chitosan treatment on enzymatic browning and antioxidant enzyme activity in sweet cherry fruit. *Postharvest Biology and Technology*, 109, 45-56.
- Petriccione, M., De Sanctis, F., Pasquariello, M. S., Mastrobuoni, F., Rega, P., Scortichini, M., & Mencarelli, F. (2015a). The effect of chitosan coating on the quality and nutraceutical traits of sweet cherry during postharvest life. *Food and Bioprocess Technology*, 8(2), 394-408.
- Petriccione, M., Mastrobuoni, F., Pasquariello, M. S., Zampella, L., ..., & Scortichini, M. (2015b). Effect of chitosan coating on the postharvest quality and antioxidant enzyme system response of strawberry fruit during cold storage. *Foods*, 4(4), 501-523.
- Poverenov, E., Danino, S., Horev, B., Granit, R., Vinokur, Y., & Rodov, V. (2014). Layer-by-layer electrostatic deposition of edible coating on fresh cut melon model: Anticipated and unexpected effects of alginate–chitosan combination. *Food and Bioprocess Technology*, 7(5), 1424-1432.

- Rahman, M., Mukta, J. A., Sabir, A. A., Gupta, D. R., Mohi-Ud-Din, M., ..., & Islam, M. T. (2018). Chitosan biopolymer promotes yield and stimulates accumulation of antioxidants in strawberry fruit. *PLOS One*, *13*(9), e0203769.
- Rastegar, S., Rahemi, M., Baghizadeh, A., & Gholami, M. (2012). Enzyme activity and biochemical changes of three date palm cultivars with different softening pattern during ripening. *Food Chemistry*, 134(3), 1279-1286.
- Reddy, M. B., Angers, P., Castaigne, F., & Arul, J. (2000). Chitosan effects on blackmold rot and pathogenic factors produced by Alternaria alternata in postharvest tomatoes. *Journal of the American Society for Horticultural Science*, 125(6), 742-747.
- Rhoades, J., & Roller, S. (2000). Antimicrobial actions of degraded and native chitosan against spoilage organisms in laboratory media and foods. *Applied and Environmental Microbiology*, 66(1), 80-86.
- Romanazzi, G., Feliziani, E., Santini, M., & Landi, L. (2013). Effectiveness of postharvest treatment with chitosan and other resistance inducers in the control of storage decay of strawberry. *Postharvest Biology and Technology*, 75, 24-27.
- Romanazzi, G., Feliziani, E., & Sivakumar, D. (2018). Chitosan, a biopolymer with triple action on postharvest decay of fruit and vegetables: Eliciting, antimicrobial and film-forming properties. *Frontiers in Microbiology*, *9*(2745).
- Ruiz-García, Y., & Gómez-Plaza, E. (2013). Elicitors: a tool for improving fruit phenolic content. *Agriculture*, *3*(1), 33-52.
- Saki, M., ValizadehKaji, B., Abbasifar, A., & Shahrjerdi, I. (2019). Effect of chitosan coating combined with thymol essential oil on physicochemical and qualitative properties of fresh fig (Ficus carica L.) fruit during cold storage. *Journal of Food Measurement and Characterization*, 13(2), 1147-1158.
- Sangsuwan, J., Pongsapakworawat, T., Bangmo, P. & Sutthasupa, S. (2016). Effect of chitosan beads incorporated with lavender or red thyme essential oils in inhibiting Botrytis cinerea and their application in strawberry packaging system. *Lebensmittel-Wissenschaft & Technologie*, 74(1), 14-20.
- Sathiyabama, M., Akila, G., & Charles, R. E. (2014). Chitosan-induced defence responses in tomato plants against early blight disease caused by Alternaria solani (Ellis and Martin) Sorauer. *Archives of Phytopathology & Plant Protection*, 47(16), 1963-1973.
- Sayyari, M., Aghdam, M. S., Salehi, F., & Ghanbari, F. (2016). Salicyloyl chitosan alleviates chilling injury and maintains antioxidant capacity of pomegranate fruits during cold storage. *Scientia Horticulturae*, 211, 110-117.

- Sayyari, M., Babalar, M., Kalantari, S., Serrano, M., & Valero, D. (2009). Effect of salicylic acid treatment on reducing chilling injury in stored pomegranates. *Postharvest Biology and Technology*, 53(3), 152-154.
- Shafiee, M., Taghavi, T. S., & Babalar, M. (2010). Addition of salicylic acid to nutrient solution combined with postharvest treatments (hot water, salicylic acid, and calcium dipping) improved postharvest fruit quality of strawberry. *Scientia horticulturae*, 124(1), 40-45.
- Shah, S., & Hashmi, M. S. (2020). Chitosan–aloe vera gel coating delays postharvest decay of mango fruit. *Horticulture, Environment, and Biotechnology*, 61(2), 279-289.
- Shen, Y., & Yang, H. (2017). Effect of preharvest chitosan-g-salicylic acid treatment on postharvest table grape quality, shelf life, and resistance to Botrytis cinereainduced spoilage. *Scientia Horticulturae*, 224, 367-373.
- Shiri, M. A., Bakhshi, D., Ghasemnezhad, M., Dadi, M., Papachatzis, A., & Kalorizou, H. (2013). Chitosan coating improves the shelf life and postharvest quality of table grape (Vitis vinifera) cultivar Shahroudi. *Turkish journal of agriculture and forestry*, 37(2), 148-156.
- Siddiq, M., Aleid, S. M., & Kader, A. A. (Eds.). (2013). *Dates: postharvest science, processing technology and health benefits* (various pages). New York: Wiley.
- Sohail, M., Ayub, M., Khalil, S. A., Zeb, A., Ullah, F., Afridi, S. R., & Ullah, R. (2015). Effect of calcium chloride treatment on post harvest quality of peach fruit during cold storage. *International Food Research Journal*, 22(6), 2225-2229.
- Terry, L. A., & Joyce, D. C. (2004). Elicitors of induced disease resistance in postharvest horticultural crops: a brief review. *Postharvest Biology and Technology*, 32(1), 1-13.
- Tian, S., Wan, Y., Qin, G., & Xu, Y. (2006). Induction of defense responses against Alternaria rot by different elicitors in harvested pear fruit. *Applied microbiology* and biotechnology, 70(6), 729-734.
- Vayalil, P. K. (2002). Antioxidant and antimutagenic properties of aqueous extract of date fruit (Phoenix dactylifera L. Arecaceae). *Journal of Agricultural and Food Chemistry*, 50(3), 610-617.
- Velickova, E., Winkelhausen, E., Kuzmanova, S., Alves, V. D., & Moldão-Martins, M. (2013). Impact of chitosan-beeswax edible coatings on the quality of fresh strawberries (Fragaria ananassa cv Camarosa) under commercial storage conditions. *LWT-Food Science and Technology*, 52(2), 80-92.

- Velioglu, Y., Mazza, G., Gao, L., & Oomah, B. D. (1998). Antioxidant activity and total phenolics in selected fruits, vegetables, and grain products. *Journal of* agricultural and Food Chemistry, 46(10), 4113-4117.
- Vyas, P. B., Tadapaneni, V. R. R., & Thakkar, V. R. (2016). Chemical elicitors improve the shelf life of phalsa (Grewia asiatica L.) by inducing antioxidants and controlling microbes. *Fruits*, 71(5), 307-317.
- Waewthongrak, W., Leelasuphakul, W., & McCollum, G. (2014). Cyclic lipopeptides from Bacillus subtilis ABS–S14 elicit defense-related gene expression in citrus fruit. *PLOS One*, 9(10), e109386.
- Wang, L., Chen, S., Kong, W., Li, S., & Archbold, D. D. (2006). Salicylic acid pretreatment alleviates chilling injury and affects the antioxidant system and heat shock proteins of peaches during cold storage. *Postharvest Biology and Technology*, 41(3), 244-251.
- Wang, S. Y., & Gao, H. (2013). Effect of chitosan-based edible coating on antioxidants, antioxidant enzyme system, and postharvest fruit quality of strawberries (Fragaria x aranassa Duch.). *LWT-Food Science and Technology*, 52(2), 71-79.
- Wang, Y. Y., Li, B. Q., Qin, G. Z., Li, L., & Tian, S. P. (2011). Defence response of tomato fruit at different maturity stages to salicylic acid and ethephon. *Scientia Horticulturae*, 129(2), 183-188.
- Xu, D., Qin, H., & Ren, D. (2018). Prolonged preservation of tangerine fruits using chitosan/montmorillonite composite coating. *Postharvest Biology and Technology*, 143, 50-57.
- Xylia, P., Chrysargyris, A., Ahmed, Z.F. R & Tzortzakis, N. (2021). Application of Rosemary and Eucalyptus Essential Oils and Their Main Component on the Preservation of Apple and Pear Fruits. *Horticulturae*, 7, 479.
- Yahia, E. M. (Ed.). (2011). Postharvest Biology and Technology of Tropical And Subtropical Fruits: Fundamental Issues (various pages). New York: Elsevier.
- Zaid, A., & De Wet, P. F. (1999). *Climatic requirements of date palm. Date palm cultivation.* FAO, Roma (Italia).
- Zeb, S., Sajid, M., Shah, S. T., Ali, M., Ali, S., Nawaz, Z., Rauf, K., Shah, A. K. H., Shah, S. A. A. & Raiz, R. (2020). 34. Influence of post-harvest application of chitosan on physico-chemical changes of apple fruit during storage. *Pure and Applied Biology (PAB)*, 9(4), 2554-2562.
- Zhang, C., Long, Y., Li, J., Li, M., Xing, D., An, H., Wu, X. & Wu, Y. (2020). A chitosan composite film sprayed before pathogen infection effectively controls postharvest soft rot in kiwifruit. *Agronomy*, 10(2), 265.

- Zhang, Y., Chen, K., Zhang, S., & Ferguson, I. (2003). The role of salicylic acid in postharvest ripening of kiwifruit. *Postharvest Biology and Technology*, 28(1), 67-74.
- Zhou, Y., Zhang, L., & Zeng, K. (2016). Efficacy of Pichia membranaefaciens combined with chitosan against Colletotrichum gloeosporioides in citrus fruits and possible modes of action. *Biological Control*, *96*, 39-47.



جامعة الإمارات العربيـة المتحدة United Arab Emirates University



UAE UNIVERSITY MASTER THESIS NO. 2022:9

This thesis is about the research conducted in the field of postharvest quality of date palm. Date fruit is the most important fruit in UAE. The research highlighted the effect of using natural compounds on extending shelf/storage life and delayed senescence, decreasing decay percentage and microbial load of 'Khesab' date fruit. These treatments when applied before harvesting were able to enhance fruit nutritional quality during long storage. The results recommended these to be used commercially on other date fruit cultivars or other fruit species to extend the fruit shelf life and improve the quality during storage.

www.uaeu.ac.ae

Shamsa Alblooshi received her MSc in Horticulture from the Department of Integrative Agriculture, College of Agriculture & Veterinary Medicine at UAE University, UAE. She received her BSc from the College of Food and Agriculture, United Arab Emirates University, United Arab Emirates in 2019.